

International Journal of Scientific Research and Reviews

Phytochemical and Antibacterial Activity on Various Extracts of *In Vivo* and *In Vitro* of *Baliospermum montanum* (Willd.) Muell. Arg.

H.R. Raveesha*¹ and B.K. Sushma²

*Department of Botany, Jnanabharathi campus, Bangalore University, Bengaluru – 560 056
Karnataka

ABSTRACT

The present study was aimed to investigate the phytochemical and antibacterial activity of *in vivo* fresh leaf and *in vitro* derived leaf callus of *Baliospermum montanum* (Willd.) Muell. Arg. An *in vitro* regeneration protocol was developed for induction of callus from leaf segments cultured on Murashige and Skoog (MS) medium supplemented with α -naphthalene acetic acid (NAA) and 6-benzyl amino purine (BAP). Phytochemical screening of major bioactive constituents of leaf and leaf derived callus of different solvents extract were analyzed. The antibacterial activity was performed by agar well diffusion method. The maximum callus induction (82.66 ± 2.51) was observed in leaf segments on MS medium supplemented with combination of BAP ($2.22 \mu\text{M}$ & $4.44 \mu\text{M/l}$) + NAA ($2.69 \mu\text{M/l}$). The phytochemical studies revealed the presence of alkaloids, flavonoids, proteins, tannins, phenols, steroids and terpenoids, phytosterols, glycosides, coumarins, carbohydrates, betacyclin, resins, gums and mucilage etc. The Methanolic extract of leaf exhibited maximum zone of inhibition against *Bacillus subtilis* (21.33 ± 1.52). Whereas acetone extract of the callus showed maximum inhibition against *Pseudomonas fluorescense* (17.0 ± 1.00) compare to other extracts. In conclusion the leaf and the callus extract reveals the presence of major phytochemicals and antimicrobial compounds and further studies has to be carried out to isolate the major bioactive compounds from *Baliospermum montanum*.

KEY WORDS: *Baliospermum montanum*, *in vitro*, callus, phytochemical, antibacterial activity.

***Corresponding Author**

Dr. H R Raveesha

Assistant Professor, Department of Botany
Bangalore University, Bengaluru - 560 056
Karnataka, India
Mobile: 919880615519
E-mail: hrraveesh74@gmail.com

INTRODUCTION

According to world health organisation (WHO) 80% of population in developing countries depend on medicinal plants for their primary health care¹. India is one of the hotspots of biodiversity for medicinal and aromatic plants in the world. Western Ghats and the North eastern region of India are the verge of extinction due to excessive plundering². Medicinal plants synthesize enormous array of secondary metabolites which are important for human life and it promote positive health and maintain resistance against infection by re-establishing body equilibrium³. A number of medicinal plants showed effective antimicrobial activities which were comparable to synthetic standard drugs⁴. Utilization of natural products as an alternative way for the convectional action in healing the different ailments has been increased in few decades⁵. An alternative source for commercial exploitation was plant cell cultures which will produce high yield compare to field grown plants⁶.

Baliospermum montanum belongs to the family Euphorbiaceae commonly known as Danti. It is a stout, monoecious under shrub with many shoots arising from the base⁷. The species is distributed from Kashmir to Arunachal Pradesh and southern peninsular India⁸. The root and leaf are rich source of metabolites like montanin, baliospermin, 12-deoxyphorbol-13-palmate, 2-deoxy-16-hydroxyphorbol-13-palmates, 12-deoxy-5 β -hydroxyphorbol-13-myristate, 8-sitosterol, 8-D-glucoside and hexacosanol. The Plant extract are used to treat as anthelmintic, diuretic, skin disease, wounds, piles, jaundice, dropsy, asthma, bronchitis and headache. Stem decoction are used to treat toothache, inflammations, flatulence and snakebite⁹⁻¹¹. To keep pace with the growing demand of this herb, evaluation and utilization of tissue culture system has an effective and alternative source to fulfill the requirements¹¹. Based on the above fact, the present study was aimed to establish effective protocol of callus induction and to investigate the phytochemical and antibacterial activity of *in vivo* fresh leaf and *in vitro* derived leaf callus.

MATERIAL AND METHODS

Collection of Plant material and sterilization:

Plant materials were collected from Western Ghats, Karnataka and maintained in green house condition. Leaves were washed thoroughly under running tap water followed by teepol and bavistin for 30 m, then in 70% ethanol for 2 m. The explants were then treated with 0.1% HgCl₂ for 2 m under aseptic condition and then washed with sterile water for 3-5 times to remove the traces of sterilants. Finally surface sterilized explants were cultured on MS medium.

Callus induction:

For callus induction, sterilized leaf segments (0.5 – 1.0 cm²) were cultured on MS medium supplemented with 3% sucrose, 0.8% agar with different concentrations and combinations of auxin (NAA) and cytokinin (BAP). Cultures were incubated at 25±2° C in a culture room with 70-80% relative humidity and 16 h of photoperiod.

Qualitative analysis:

5g of fresh leaf and callus were soaked in different solvents namely aqueous, methanol, ethanol, acetone, butanol, chloroform, petroleum ether and hexane for 3-4 days. The solvent extracts were filtered with Whatman filter paper No.1. The procedure was repeated for another two cycles to ensure complete extraction of phytochemical compounds and the filtrates were stored at 4° C until the further analysis¹². The lyophilized extracts were dissolved in respective solvents and were screened for the qualitative analysis for the presence of alkaloids, flavonoids, proteins, tannins, phenols, steroids and terpenoids, phytosterols, glycosides, coumarins etc., by standard methods¹³⁻¹⁶.

Antibacterial assay:

Antibacterial activity was performed by well diffusion method following the protocol of Johnson *et al.*⁹. The bacterial strains *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, *Klebsiella pneumoniae* and *Pseudomonas fluorescense* were obtained from Department of Microbiology, Bangalore University, Bengaluru. Nutrient agar was prepared using beef extract 3 g/l, peptone 5 g/l, sodium chloride 5 g/l and agar 15 g/l with pH adjusted to 7.2. Fresh leaf and callus were subjected for antibacterial activity against pathogenic bacteria. Petri plates were prepared with nutrient agar media and wells were bored with 8mm diameter sterilized cork borer. Different solvent extracts were used for bacterial sensitivity test. Plates were incubated for 24 h at 37° C and zone of inhibition were recorded in mm and compared with standard antibiotics (Streptomycin and Tetracyclin).

Statistical Analysis:

The data are expressed as Mean ± S.E. All the experiments were repeated thrice, data were analysed statistically by one way analysis of variance followed by Duncan's multiple range tests using SPSS software. Probability values $P < 0.05$ were considered significant.

RESULTS AND DISCUSSION:

Callus induction:

Growth hormones play an important role in tissue culture for the development of cultured cells or tissues¹⁷. It also regulates the various physiological and morphological processes and enhances metabolites synthesis^{18,19}. In the present study, different concentration of cytokinins and auxins were supplemented for the callus induction. The green friable callus was observed at the cut ends of leaf explants on MS medium supplemented with different concentrations and combinations of BAP (2.22 μ M-17.75 μ M/l) and NAA (2.69 μ M-21.48 μ M/l) (Fig.2). Callus induction was maximum in MS medium supplemented with BAP (2.22 μ M & 4.44 μ M/l) + NAA (2.69 μ M/l) after six weeks of inoculation (Fig. 1). The competence of callus induction depends on the type of growth regulators, explants source and culture medium²⁰. Previous studies by Teshome *et al.*²¹ and Kumari *et al.*²² reported higher callus induction from leaf explants inoculated on MS medium supplemented with NAA and BAP.

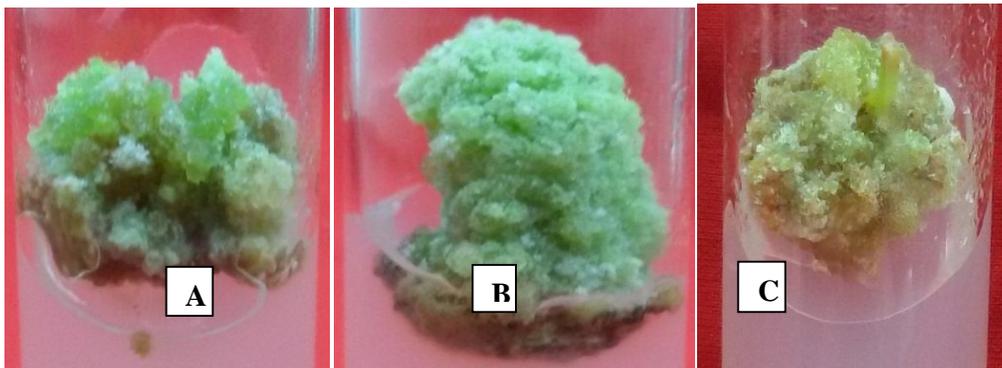


Fig. 1: A-C Induction of semi friable green callus from leaf explants

A: MS+BAP (2.22 μ M)+NAA (2.69 μ M), B: MS+BAP (4.44 μ M) +NAA (2.69 μ M) and C: MS+BAP (17.75 μ M) +NAA (2.69 μ M)

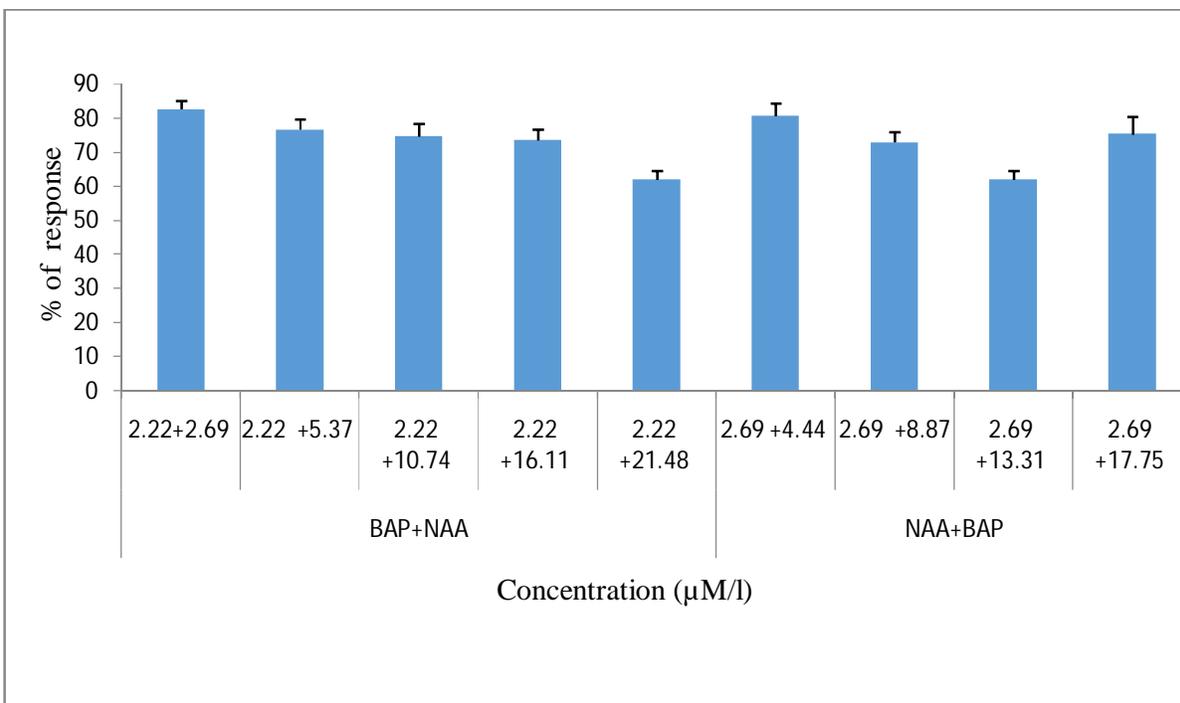


Fig. 2: Percentage of callus induction on different concentrations and combinations of auxin and cytokinin of *B. montanum* from leaf explants.

Phytochemical analysis:

Medicinal plants were found to contain various phytochemical produced at different stage of development under stress condition²³. Secondary metabolites has a biological and therapeutic properties and it serves to protect the plant growth against microbial attacks and from animal predators. Qualitative phytochemical analyses of different solvent extracts were performed in leaf and callus of *B. montanum*. Our results indicated the presence of alkaloids, flavonoids, proteins, tannins, phenols, steroids and terpenoids, phytosterols, glycosides, carbohydrates, betacyanin, resins, Phlobatannins, volatile oils, and coumarins (Table.1). Methanolic extract proved to be a better solvent compared to other extracts. Sangha *et al.*²³ reported that methanolic extract is a good solvent to extract major phytochemicals from root, stem, leaf, latex and flower extracts in *B. montanum*. Studies by different workers demonstrated that methanolic extract proves to be a better solvent for the isolation of phytochemicals from different plant species²⁴⁻²⁶.

Table.1: qualitative analysis of *B. montanum* fresh leaf and fresh callus

TESTS	FRESH LEAF								FRESH CALLUS							
	I	II	III	IV	V	VI	VII	VIII	I	II	III	IV	V	VI	VII	VIII
Alkaloids	-	-	-	+	+	-	+	-	-	-	+	-	+	-	-	+
Flavonoids	+	+	+	-	+	-	+	-	+	+	+	-	-	-	+	+
Proteins	+	+	+	+	+	+	+	+	+	+	+	-	-	+	+	+
Phenols	+	+	+	-	-	-	+	-	+	+	+	-	-	-	-	+
Tannins	+	+	+	-	-	-	+	-	+	+	+	-	-	-	-	+
Steroids &Terpenoids	+	+	+	+	-	+	+	+	+	-	-	-	-	-	+	-
Phytosterols	+	+	+	+	-	+	+	+	+	-	-	-	-	-	+	-
Glycosides	+	+	+	-	+	-	+	+	+	+	+	-	-	-	+	+
Coumarins	+	+	+	+	+	-	-	+	-	-	-	+	+	+	-	+
Carbohydrates	+	+	+	-	+	-	+	+	+	+	+	+	+	+	+	+
Betacyanin	+	+	+	+	+	-	-	+	+	+	+	+	-	-	-	+
Resins	-	+	+	-	-	-	-	-	+	+	+	+	-	+	-	-
Phlobatannins	+	+	+	-	-	-	-	+	+	+	+	-	-	-	-	-
Starch	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Volatile oils	-	+	+	-	-	-	-	-	+	+	+	-	-	-	-	+
Emodols	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-
Gums and mucilage	-	+	+	-	+	-	-	+	-	-	-	-	-	-	-	-

I-Aqueous, II-Ethanol, III-Methanol, IV-Petroleum ether, V-Chloroform, VI-Hexane, VII-Butanol, VIII-Acetone

Antibacterial activity:

Plant extracts can be good source of antibiotics against various fungal and bacterial pathogens. Plant based antimicrobial compounds have enormous therapeutically potential and they have no side effect compare to synthetic compounds²⁷. The results of antibacterial activity of different solvents extracts of leaf and leaf derived callus were represented (Fig. 3 & 4). The result revealed that methanolic and aqueous leaf extract showed maximum activity against *Bacillus subtilis* (21.33±1.527) and *Klebsiella pneumoniae* (21.00±1.00). Whereas in callus, acetone and ethanolic extracts showed higher inhibitory activity against *Staphylococcus aureus* (17.00±1.00) and *Escherichia coli* (16.66±1.52). The results are concordance with the report of Johnson *et al.*⁹ wherein ethanolic extract of *B. montanum* (leaf, root and callus) showed maximum inhibition against different bacterial pathogen.

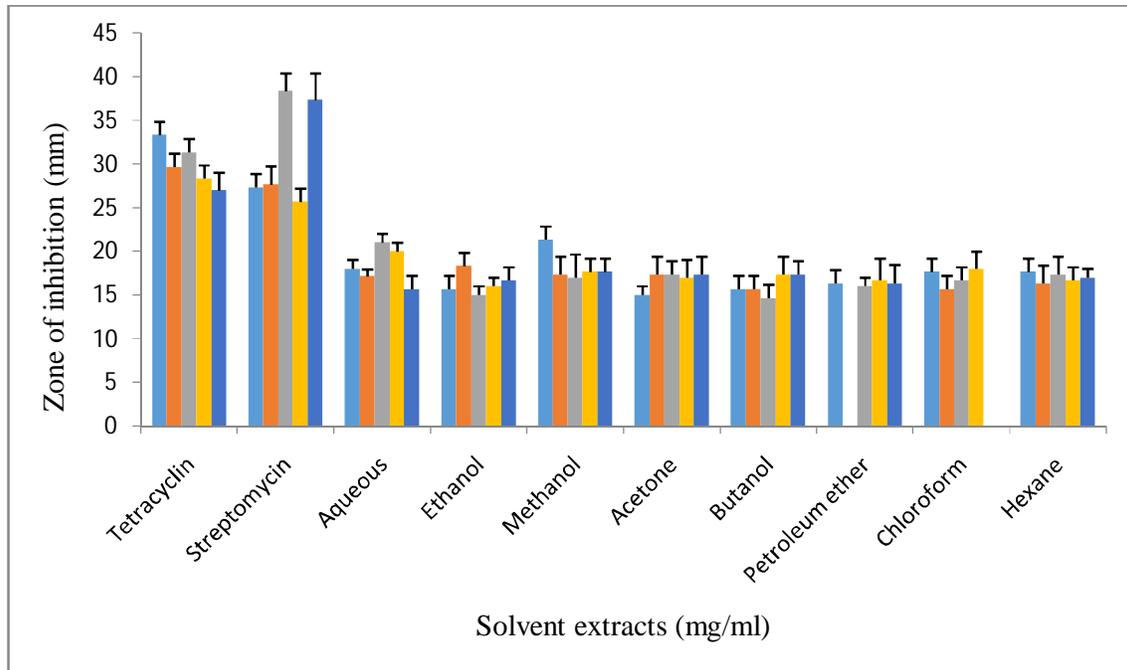


Fig. 3: Zone of inhibition in different solvents of *B. montanum* from fresh leaf

B. subtilis *E. coli* *K. Pneumonia* *P. Fluorescence* *S. aureus*

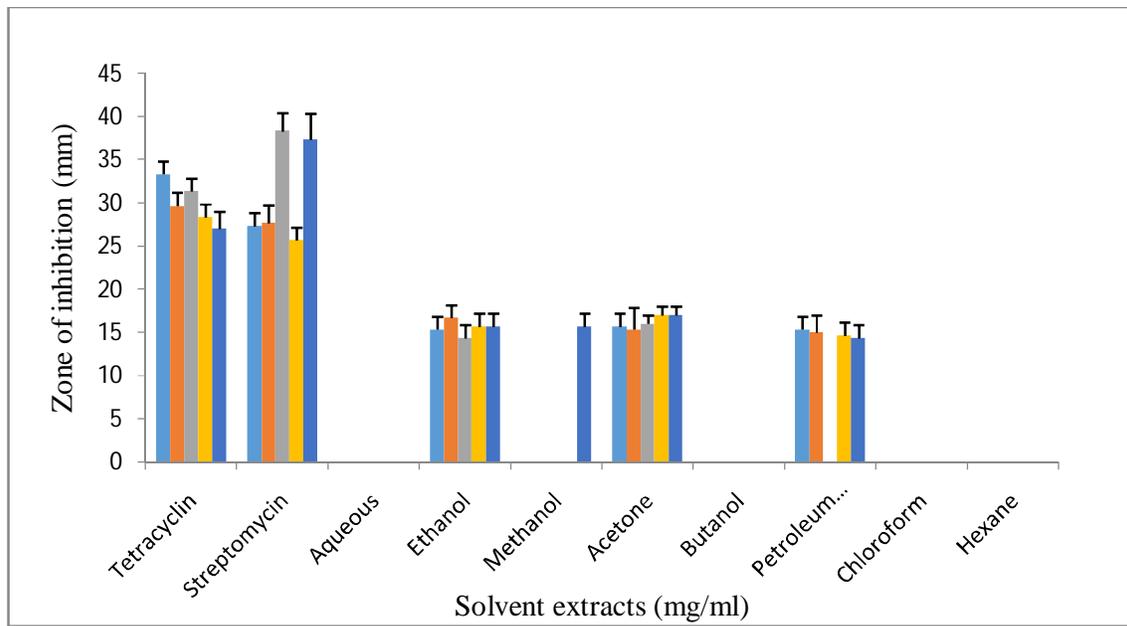


Fig. 4: Zone of inhibition in different solvents of *B. montanum* from fresh callus

B. subtilis *E. coli* *K. pneumoniae* *P. fluorescense* *S. aureus*

CONCLUSION:

In conclusion, maximum callus induction was noticed in lower concentration and combination of NAA and BAP. Methanol and acetone are found to be a better solvent for the extraction of major phytochemical from *B. montanum*. The methanol and acetone of leaf and callus were found to be more effective against pathogens compared to other extracts. Further studies have to be carried out on the isolation and identification of antimicrobial compounds.

ACKNOWLEDGEMENTS:

The authors are thankful to Department of Microbiology and Biotechnology, Bangalore University for providing the Bacterial strains.

REFERENCES:

1. Vines G. Herbal harvest with future: towards sustainable source for medicinal plants. Plant life international www.plantlife.co.uk 2004.
2. Gupta V. Seed germination and dormancy breaking techniques for indigenous medicinal and aromatic plants. J. Med. Aromatic Pl. Sci. 2003; 25: 402-407.
3. Savant C, Joshi N, Reddy S, Mannasaheb BA, Joshi H. Immuno modulatory medicinal plants of India: A Review. Int. J. Pharma. Toxicol. 2014; 4: 109-115.
4. Naz R, Bano A. Antimicrobial potential of *Ricinus communis* leaf extracts in different solvents against pathogenic bacterial and fungal strains. Asian Pacific J. Tro. Bio. 2012; 2: 944-947.
5. Saeed S, Tariq P. Antimicrobial activities of *Embllica officinalis* and *Coriandrum sativum* against Gram-positive bacteria and *Candida albicans*. Pak. J Bot. 2007; 39: 913-917.
6. Shilpa K, Varun K, Lakshmi B. An alternative method of natural drug production: eliciting secondary metabolite production using plant cell culture. J. Pl. Sci. 2010; 5: 222-247.
7. Kumar SV, Mishra SH. Protective effect of extracts of *Baliospermum montanum* (Wild.) Muell. Arg. against paracetamol-induced hepatotoxicity-an *in vivo* and *in vitro* study. Ancient Sci. Life. 2015; 33: 216-221.
8. Chopra RN, Nayar SL, Chopra IC. Glossary of Indian Medicinal Plants New Delhi. Council of Scientific and Industrial Research. 1956.
9. Johnson M, Wesely EG, Hussain Z, Selvan N. *In vivo* and *in vitro* phytochemical and antibacterial efficacy of *Baliospermum montanum* (Willd.) Muell. Arg. Asian Pacific J. Tro. Med. 2010; 894-897.

10. Sivarajan VV, Balachandran. Ayurvedic Drugs and their Plant Sciences. Oxford and IBHI Publishing Co.Pvt.Ltd: New Delhi; 1994; 125-126.
11. Bourgaud F, Nguyen C, Guckert A. *In vitro* culture and production of furanocoumarins and secondary metabolites. *Berlin Heidelberg: Springer-Verlag*.1995; 388-411.
12. Dhanapal ACTN, Ming TW, Aung HP, Hao SJ. Preliminary screening of *Artemisia argyi* for antioxidant potentials. *Int. J. Pharmacog. Phytochem. Res.* 2016; 8: 347-355
13. Harborne JB. Phytochemical methods. A guide to modern technique of plant analysis, 2nd ed. Chapman: 1984; 100-101.
14. Kokate CK. Practical Pharmacognosy. Vallabh Prakashan, New Delhi. 1994; 1: 15-30.
15. Sofowora A. Medicinal Plants and Traditional Medicine in Africa. Spectrum Books.Ltd.(Pub.): Ibadan; 1993; 1-153.
16. Raman N. Phytochemical Techniques. New Indian Publishing Agencies; 2006; 19.
17. Ngomuo M, Mneney E, Ndakidemi P. The effects of auxins and cytokinin on growth and development of (*Musa* sp.) Var. “Yangambi” explants in tissue culture. *American J. Pl. Sci.* 2013; 4: 2174-2180.
18. Rayns FW, Fowler MR, Hunter CF. Media design and use *in vitro* cultivation of plant cells. Butterworth – Heinemann Ltd: Oxford; 1993; 43-64.
19. Srivastava LM. Plant growth and development. Hormones and Environmental Academic Press: New York; 2002; 140-143.
20. Sen MK, Nasrin S, Rahman S, Mostofa AH. *In vitro* callus induction and plantlet regeneration of *Achyranthus aspera* L., a high value medicinal plant. *Asian Pacific J. Bio.* 2014; 4: 40-46.
21. Teshome S, Feyissa T. *In vitro* callus induction and shoot regeneration from leaf explants of *Glinus lotoides* (L.) - An important medicinal plant. *Asian J. Pl. Sci.* 2015; 6: 1329-1340.
22. Kumari M. Evaluation of methanolic extracts of *in vitro* grown *Tinospora cordifolia* (Willd.) for antibacterial activities. *Asian J. Pharm. Cli. Res.* 2012; 5: 172-175.
23. Sangha RB, Gayathri MC, Rajanna L. Phytochemical profile of *Baliospermum montanum* (Wild.) Muell. Arg. *Int. J. Innove. Sci. Res.* 2014; 9: 54-60.
24. Senguttuvan J, Paulsamy S, Karthika K. Phytochemical analysis and evaluation of leaf and root parts of the medicinal herb, *Hypochoeris radicata* L. for *in vitro* antioxidant activities. *Asian Pacific J. Bio.* 2014; 4(Suppl 1): 5359-5367.
25. Anupa MP, Vasanthakumari B, Citarasu T. Qualitative phytochemical screening of leaf extracts of *Cissampelos pareira* L. in different solvent systems. 2014; 3399-3403.

26. Gopinath SM, Rakesh CK, Ashwini GM, Dayananda KS. Preliminary phytochemical evaluation of leaf extracts of *Euphorbia hirta*, *Syzygium cumini* of Siddarabetta, Tumkur district, Karnataka. *Int. J. Pharma.Biosci.* 2012; 3: 431-434.
 27. Shariff N, Sudarshana MS, Umesha S, Hariprasad P. Antimicrobial activity of *Rauvolfia tetraphylla* and *Physalis minima* leaf and callus extracts. *African J. Biotech.* 2006; 5: 946-950.
-