

International Journal of Scientific Research and Reviews

Green Synthesis of Silver Nanoparticles Using *Passiflora foetida* Extract and Screening for Their Catalytic Activity

**Vundela Reddy Swetha¹, Bonigala Bodaiah¹, Mangamuri Kiranmayi Usha²,
Poda Sudhakar¹, Kondapalli Kasturi^{1*}**

¹Department of Biotechnology, Acharya Nagarjuna University, Guntur,
Andhra Pradesh, India - 522510

²Department of Botany and Microbiology, Acharya Nagarjuna University, Guntur,
Andhra Pradesh, India - 522510

ABSTRACT

Green synthesis is a zone of biosynthesis in which metallic nanoparticles are formulated using the extracts of plants as reducing and stabilizing agents. In the present study silver nanoparticles (AgNPs) were green synthesized using the aqueous extract of *Passiflora foetida*. When the plant extract was added to AgNO₃ solution, the amalgamated solution turned deep reddish brown in colour after 48 hours indicating the formation of AgNPs. Later in UV-Visible analysis the amalgamated solution has shown absorption maximum at 435.13nm which confirmed the formation of AgNPs. The green synthesized AgNPs were further characterized using dynamic light scattering, zeta potential analysis, X-ray diffractometer (XRD) and Transmission electron microscopy (TEM) to know their size, stability and crystalline nature. The plant mediated AgNPs were studied in fluorescence transmission infrared spectroscopy (FTIR) to know the phytochemicals present in the plant extract which acted as reducing agents in the formation of AgNPs. Imperatively in the present study the plant mediated AgNPs were screened for catalytic activity in the presence of NaBH₄ in the degradation and removal of 4-Nitrophenol, methylene blue, methyl orange and methyl red. The obtained results proclaimed that the AgNPs of *P. foetida* acted as potential catalyzing agents in the degradation of 4-Nitrophenol, methylene blue, methyl orange and methyl red.

KEYWORDS: Green synthesis, *Passiflora foetida*, AgNPs, XRD, FTIR, Catalytic activity

***Corresponding author**

Dr. Kondapalli Kasturi

Assistant Professor

Department of Biotechnology, Acharya Nagarjuna University

Nagarjunanagar, Guntur, Andhra Pradesh, India-522510

Email: kasturi.is.kondapalli21@gmail.com

Land line: 0866 -2494277, Mobile no: 9848546664

INTRODUCTION

Water pollution is a major issue in the present century because huge amounts of industrial effluents with remnant toxic chemicals are transported or thrown in to water bodies. The toxic pollutants have shown negative impact on human health and other living organisms¹. Waste water treatment by physical and chemical methods requires high cost equipment which consume huge amount of energy. There is an urgent need to develop a new technology which can replace the physical and chemical methods for the removal of water pollution. According to research records metallic nanoparticles (MNPs) especially biosynthesized MNPs in addition to their potent biological activities have shown remarkable catalytic activity in the area of degradation and removal of toxic chemicals (for example 4-Nitrophenol) and synthetic dyes^{2, 3}. Green synthesis is a sector of biosynthesis in which MNPs are synthesized in large scale using the extracts of plants in cost effective and ecofriendly manner. Among MNPs, silver nanoparticles (AgNPs) received high acknowledgements from the scientific world because of their remarkable antimicrobial, anticancer, insecticidal and catalytic activities etc⁴.

4-Nitrophenol is a toxic synthetic chemical and polymorphic compound in crystalline state. It is colorless below pH 5.4 and yellow above pH 7.5 in solution form. 4-Nitrophenol is used to darken the leather and in the manufacture of drugs, fungicides, insecticides and dyes. Ingestion and inhalation of 4-Nitrophenol leads to nausea, drowsiness, headache and cyanosis⁵. Dyes are the major synthetic organic compounds used in textile industries for coloration of the fabrics. After completing the process i.e coloration of fabrics 15% of dye is wasted and released in to water bodies which ultimately results in to serious water pollution. Methylene blue or methylthioninium chloride is a synthetic heterocyclic aromatic dye and dark green powder when dissolved in water turns in to blue colour. It is a safe drug for therapeutic use in lower doses. Methylene blue has shown antimalarial activity and increases arterial pressure in septic shock. When exposed to higher doses it causes head ache, fever, dizziness and mental confusion⁶. Methyl orange is a synthetic dye, pH indicator in acid-base titrations and causes irritation incase of skin contact and eye contact. Another one methyl red is an azodye and used in acid base titrations as pH indicator. It is prepared by diazotization of anthranilic acid and followed by dimethyl aniline⁷.

Passiflora foetida is a native plant of Southwestern United states and Mexico, belongs to the family Passifloraceae and has been introduced to the tropical regions of the world. The plant stem is thin and wiry with minute yellow hairs in younger stage and becomes woody later. The leaves are three to five lobed and the flowers are white to pale cream colored. The fruits of the plant are globose, 2-3cm in diameter and when riped appear yellow-orange to red in colour. The aqueous and

ethanolic extracts of plant has shown the presence of various secondary metabolites like terpenoids, phenols, saponins, flavanoids, steroids, alkaloids, oils and cardioglycocides^{8, 9}. In the present study AgNPs were synthesized using the aqueous plant extract of *P. foetida* and characterized using different advanced techniques. Further the green synthesized AgNPs were screened for their catalytic activity in the degradation and removal of 4-Nitrophenol, Methylene blue, Methyl orange and methyl red.

MATERIALS AND METHODS

Collection of the plant material

The aerial parts of *Passiflora foetida* were collected from Tirumala hills, Tirupathi, Andhra Pradesh, India. The plants were taxonomically identified and authenticated by Prof. M. Vijayalakshmi, Department of Botany and Microbiology, Acharya Nagarjuna University, Guntur, Andhra Pradesh, India.

Preparation of the aqueous extract of P. foetida

The plant material collected was washed thrice with distilled water to remove the dust and dried under the shade to remove the moisture. The dried plant material was then cut in to pieces and crushed in to fine powder with a suitable pulveriser. 3 grams of finely crushed dried powder was mixed to 100 ml of molecular grade water, boiled at 100⁰C for 10 minutes and the extract was filtered with Whatman No 1 filter paper to remove impurities.

Green synthesis of AgNPs from the extract of P. foetida

2ml, 10ml, 20ml, 30ml and 40ml of filtered plant extract was added to 198ml, 190ml, 180ml, 170ml and 160ml of 1 mM Silver nitrate (AgNO₃) solution respectively and kept for incubation. The effect of AgNO₃ concentration on AgNPs formation was analyzed by adding 10ml of plant extract to 190ml of 0.1mM, 0.5mM, 1mM, 1.5mM and 2mM concentrations of AgNO₃ in separate reactions. Later the suspension was kept for incubation at room temperature.

Characterization of plant mediated AgNPs

The formation of AgNPs using *P. foetida* extract was confirmed by UV-Visible spectroscopic studies after 48hrs using AgNO₃ solution as blank and the values were recorded within the range of 200 to 800 nm. To know the effect of time on AgNPs formation, the amalgamated solution containing 10ml of plant extract + 190ml of AgNO₃ of AgNO₃ was analyzed using UV-

Visible spectrometer for every 1hr time intervals. Later the plant mediated AgNPs were examined in HORIBA particle size analyzer and zeta potential analyzer to know the particle size and stability of AgNPs respectively.

The AgNPs were purified by repeated centrifugation at 10,000 rpm for 15 min. The pellet of AgNPs were transferred into china dish and kept for shade evaporation. The dried nanoparticles were washed with distilled water, allowed for shade drying and the process was repeated thrice. The purified and dried nanoparticle samples were collected and used for further characterization. The purified AgNPs were studied using Philips X'pert pro XRD with an operation voltage of 40KV and current of 30mA with CuK α radiation (1.540 Å) between 2 θ angles (30°-80°) for analysing peak data and crystal structure. Fluorescence transmission infrared (FTIR) analysis of the AgNPs was carried out through potassium bromide (KBr) pellet (FTIR grade) method in 1:100 ratio and spectrum was recorded using Jasco FT/ IR- 6300 FTIR equipped with JASCO IRT-7000 Intron Infrared microscope (JASCO, Tokyo, Japan) using transmittance mode operating at a resolution of 4cm⁻¹ in order to find out the phytochemicals in *P. foetida* extract which are responsible for reduction process in the AgNPs synthesis. Transmission electron microscope study was carried out to know the morphology and particle size distribution of AgNPs. The grid for TEM analysis was prepared by placing a drop of *P. foetida* mediated AgNPs suspension on a carbon-coated copper grid and allowing the water to evaporate inside a vacuum dryer. The grid containing AgNPs was scanned by a Hitachi Japan Model 7500 TEM machine.

Evaluation of catalytic activity of plant mediated AgNPs

In the present study AgNPs of *P. foetida* were utilised as catalysts separately in the degradation and removal of 4-Nitrophenol, Methylene blue, Methyl orange and Methyl red by NaBH₄. The procedure of degradation of a respective synthetic chemical or dye (4-Nitrophenol or Methylene blue or Methyl orange or Methyl red) by NaBH₄ in presence of AgNPs as catalyst involves 3 reactions. All the reactions were studied in Thermoscientific UV-Visible spectrophotometer using milli Q water as blank. The first reaction is prepared by adding 1.5mL of 1mM of a synthetic chemical or dye to 1.5mL of milli Q water, mixed well and analyzed in UV-Visible spectroscopy. Later 1mg of solid NaBH₄ was added to first reaction to prepare second reaction and analyzed in UV-Visible spectroscopy. The third reaction is prepared by adding 10 μ L of green synthesized nanosolution to the second reaction and analyzed after 1 minute in UV-Visible spectroscopy⁷.

RESULTS AND DISCUSSION

Addition of *P. foetida* plant extract with aqueous solution of silver nitrate led to observable Colour change from yellowish to dark reddish brown solution after 48 hrs incubation due to Surface Plasmon Resonance indicating AgNPs formation¹⁰ and the result was depicted in the Fig.1d.

Characterization of *Passiflora foetida* AgNPs

UV-Visible analysis

The amalgamated solution of 190ml AgNO₃ + 10ml plant extract have shown absorption maximum at 435.13 (Fig. 2c) in UV-Visible spectroscopic analysis after 48 hrs incubation^{11, 12} which confirmed the formation of AgNPs. When the amalgamated solutions that kept for incubation to know the effect of plant extract concentration studied in UV-Visible spectroscopy AgNPs formation was confirmed in the reactions with 2ml, 10ml and 20ml of plant extract only (Fig. 2a). Further when the amalgamated solutions that kept for incubation to know the effect of AgNO₃ concentration studied in UV-Visible spectroscopy AgNPs formation was confirmed in the reactions with only to 1mM, 1.5mM and 2mM concentrations of AgNO₃ (Fig. 2b). In another study when the amalgamated solution of 190ml AgNO₃ + 10ml plant extract was studied every 1hr time intervals in UV-Visible spectroscopy *P. foetida* AgNPs formation was observed after 27hrs (Fig. 2c).

Dynamic light scattering (DLS) study

In DLS study the particles exhibit Brownian motion when dispersed in the medium which is measured by the fluctuations in the intensity of scattered light in the system from which translational diffusion co-efficient is calculated by applying Stokes-Einstein equation which determines the hydrodynamic size¹³. The graph has shown almost equal size distribution of *P. foetida* AgNPs and their mean size was recorded as 99.3nm (Fig. 3a). Polydispersity index (PDI) represents the ratio between different sizes to total number of particles. A PDI value more than 0.5 refers to the aggregation of the particles¹⁴. The green synthesized AgNPs have shown zero PDI value which clearly indicates that the particles are in mono dispersed phase with no aggregation.

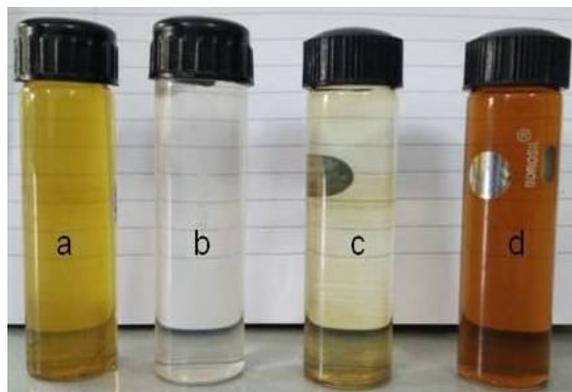


Fig. 1 Green synthesis of AgNPs (a) Aqueous plant extract (b) Silver nitrate solution (c) 1mM silver nitrate+plant extract at the start of incubation (d) *P. foetida* AgNPs

Zeta potential analysis

The Zeta potential of *P. foetida* AgNPs solution is measured by applying an electric field across the dispersion. Particles within the dispersion with a zeta potential will migrate towards the electrode of opposite charge with a velocity proportional to the magnitude of zeta potential. It should be noted that the particles with zeta potential values more positive than +30 mV or more negative than -30 mV are considered to be stable¹⁵. The zeta potential of *P. foetida* AgNPs was recorded as -20.8mV in the zeta potential analysis as shown in the Fig. 3b and from the obtained result it can be known that the synthesized nanoparticles are moderately stable¹⁶.

XRD Analysis

X-ray powder diffraction spectrum of green synthesized AgNPs (Fig. 4a) have shown Bragg peaks (angle 2θ) at 27.48° , 31.90° , 37.78° , 43.80° , 45.91° , 57.15° , 64.25° and 76.56° which corresponds to the indexed planes of 210, 122, 111, 200, 142, 241, 220 and 311 miller indices of face centered cubic (FCC) structure of a regular silver crystal¹⁷. Using Debye -Scherrer equation the average particle size of biosynthesized was determined [$d = K\lambda / \beta \cos \theta$] where 'd' is the mean diameter of the particle; 'K' is the shape factor (0.9); ' λ ' is the X-ray radiation source (0.154 nm); ' β ' is $(\pi / 180) * \text{FWHM}$ and ' θ ' is the Bragg angle. The average particle size of *P. foetida* AgNPs was obtained as 14.30nm and the XRD pattern was in agreement with earlier XRD reports of green synthesized AgNPs¹⁸.

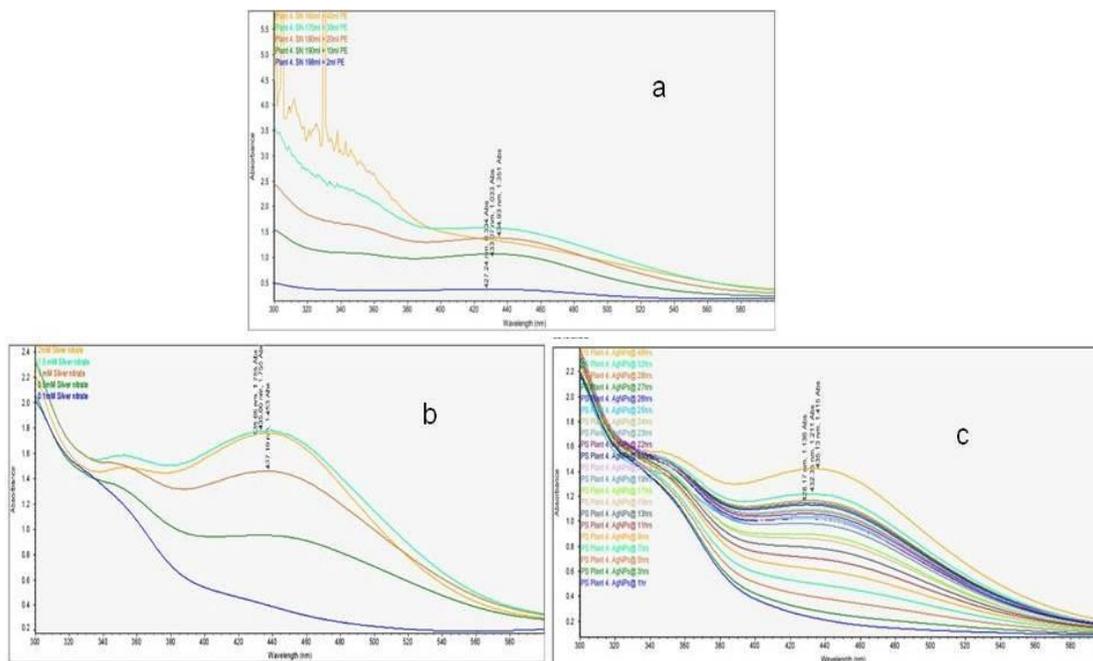


Fig. 2 UV-Visible analyses of (a) Effect of plant extract concentration (b) Effect of AgNO_3 concentration (c) Influence of time on *P. foetida* AgNPs formation

FTIR study

The biosynthesized AgNPs displayed a number of peaks in the FTIR spectrum (Fig. 4b) and portrayed the complex nature of particles. The strong and broad peak at 3371 cm^{-1} was formed because of O-H bond stretching of alcohols and phenols¹⁹. The peaks formed at 2923 cm^{-1} and 2854 cm^{-1} due to the characteristic stretching vibrations of C-H stretch of alkanes. The medium and strong peak at the 1611 cm^{-1} is the N-H stretching vibrations of Amides¹⁸. The medium peak at 1382 cm^{-1} was formed because of $-(\text{CH}_3)_3$ bend of alkanes and alkyl groups. The strong peak formed at 1222 cm^{-1} due the stretching vibrations of alkyl groups. The peak at 1055 cm^{-1} was formed due to the C-O stretch of alcohols. A strong peak was formed at 986 cm^{-1} because of the $=\text{C-H}$ bend of alkenes. The C-H bend of aromatic compounds formed a weak peak at 708 cm^{-1} . These shifts in peak positions revealed the presence of different secondary metabolites in the plant extract of *P. foetida*. Hence it can be concluded that the phytochemicals or secondary metabolites of plant extract acted as reducing and stabilizing agents in the AgNPs formation⁵.

TEM analysis

The TEM image revealed that most of the AgNPs were spherical in shape with an average size of 20nm (Fig. 4c) and is in agreement with results produced in XRD analysis²⁰. The TEM examination also confirmed the presence of biomolecule blanket on the surface of AgNPs which acts

as capping agents. Therefore the AgNPs of *P. foetida* were polydispersed without direct contact with the other and stable for longer periods of time.

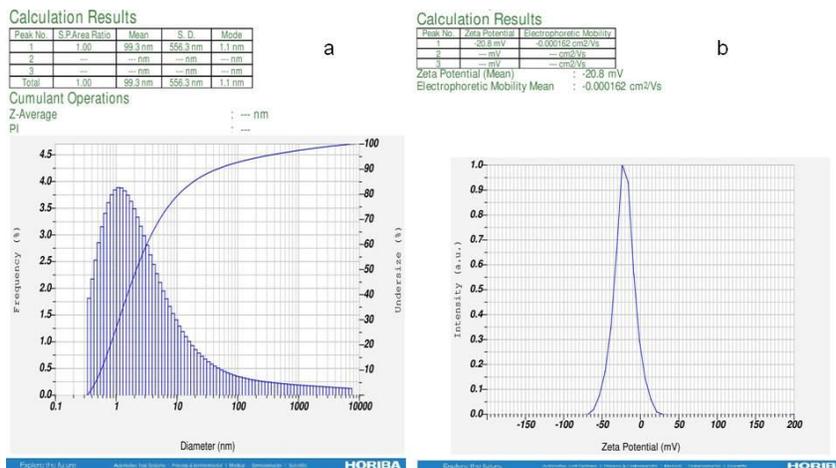


Fig. 3 (a) Dynamic light scattering (b) Zeta potential analysis of *P. foetida* AgNPs

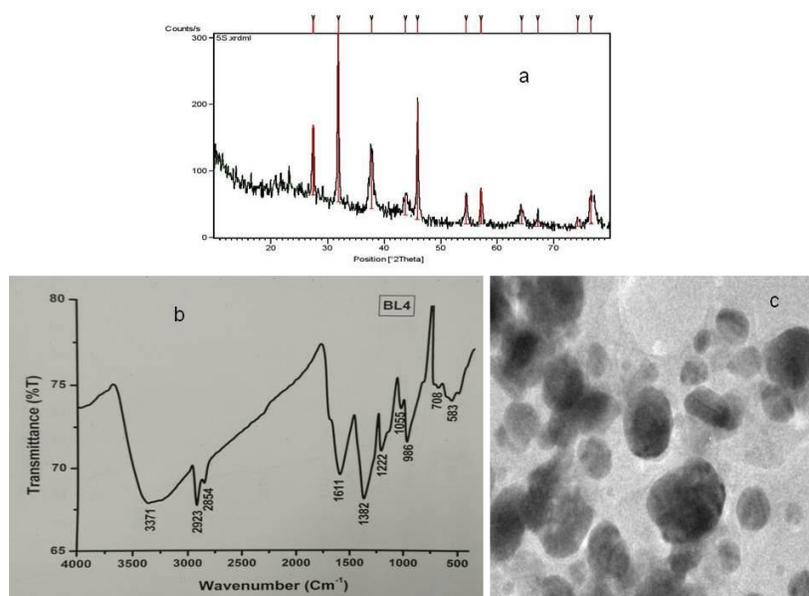


Fig. 4 (a) XRD spectrum (b) FTIR spectrum (c) TEM image of *P. foetida* AgNPs

Catalytic activity of *P. foetida* AgNPs

The synthetic chemical or dye degradation reactions were monitored and depicted in the following order 4-Nitrophenol, Methylene blue, Methyl orange and Methyl red. All reactions were analysed by UV-Visible spectrophotometer.

Reduction reactions of 4-Nitrophenol

UV-Visible analysis of 4-Nitrophenol degradation using NaBH₄ with *P. foetida* AgNPs as catalysts were shown in the Fig. 5a. The reaction of 4-Nitrophenol when monitored in

spectrophotometer the absorption maximum (1.252) was observed at 318 nm. On addition of NaBH₄ to first reaction the solution appeared bright yellow in colour because of the formation of sodium phenolate, no change in the absorption maximum with respect to time was observed and shifted to 400 nm²¹. With the addition of plant mediated AgNPs the solution turned colourless and the absorption maximum was suddenly decreased from 1.252 to 0.823 indicating the complete degradation of 4-Nitrophenol²².

Reduction reactions of Methylene blue

The degradation and removal of methylene blue by NaBH₄ in the presence of biosynthesized AgNPs as catalyst were analyzed and the UV-Visible results were illustrated in Fig. 5b. For pure 1mM methylene blue absorption maximum of 2.009 was initially observed at 664 nm²³. When 1mg of NaBH₄ added the absorption maximum was recorded at same wavelength with a slight change. With the addition of green synthesized AgNPs the solution turned colourless and the absorption maximum was completely decreased from 2.009 to Nil indicating that methylene blue was completely degraded²⁴.

Reduction reactions of Methyl orange

Fig. 5c illustrates the results related to the degradation and removal reactions of methyl orange by NaBH₄ in the presence of *P. foetida* AgNPs as catalysts. In UV-Visible analysis for pure methyl orange the absorption maximum was found to be 0.752 at 462.22nm and when 1mg of NaBH₄ added the absorption maximum was found to be 1.673 at 461.11nm. After adding AgNPs, the solution turned colourless with decrease in absorption maximum from 1.673 to 0.647²⁵. From the above result it is clearly known that the AgNPs of *P. foetida* completely degraded methyl orange in presence of NaBH₄.

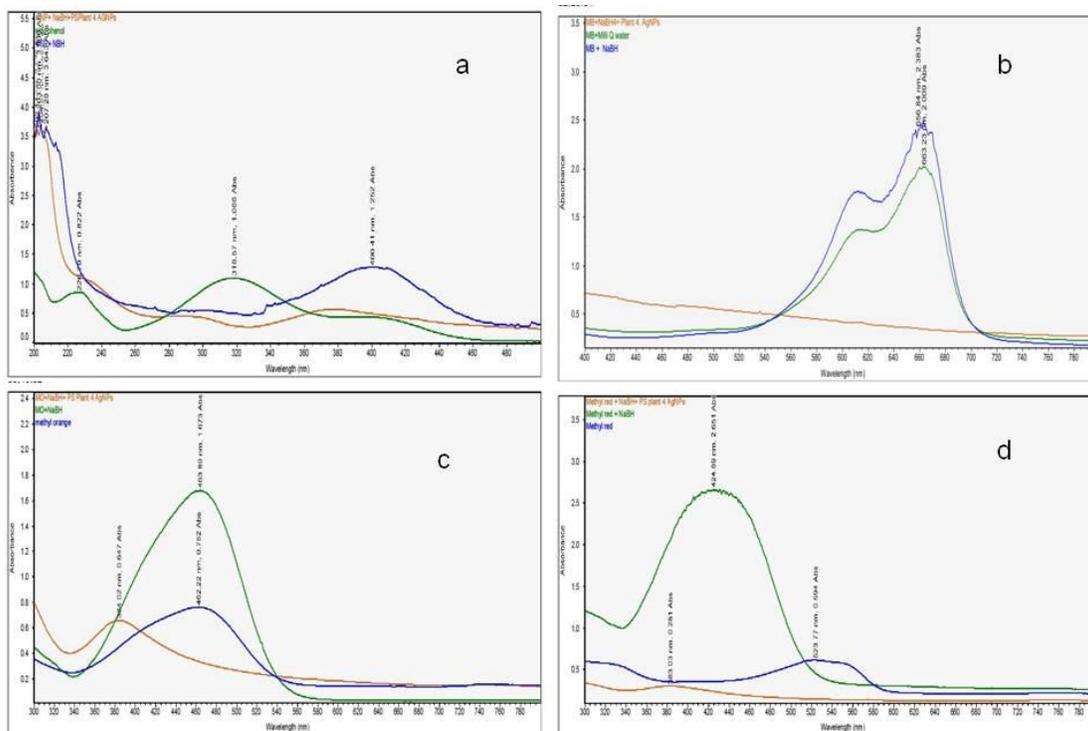


Fig. 5 Catalytic activity of *P. foetida* AgNPs on (a) 4-Nitrophenol (b) Methylene blue (c) Methyl orange (d) Methyl red (NaBH: Sodium tetra borate, AgNPs: Silver nanoparticles)

Table No. 1- Absorption maxima of synthetic chemicals before and after addition of AgNPs

Name of the synthetic chemical/ dye	Absorption maximum before the addition of AgNPs	Absorption maximum after the addition of AgNPs
4-Nitrophenol	1.252	0.822
Methylene blue	2.009	Nil
Methyl orange	1.673	0.647
Methyl red	2.651	0.281

Reduction reactions of Methyl red

UV-Visible analysis results related to the reduction reactions of methyl red by NaBH₄ in presence of AgNPs as catalysts were illustrated in Fig. 5d. An absorption maximum of 0.594 was observed at 523.77nm to pure 1mM methyl red. After addition of 1mg NaBH₄ the absorption maximum of 2.651was obtained at 424.89nm. With the addition of *P. foetida* AgNPs yellow colour

solution was turned colourless and absorption maximum of 0.281 was recorded for third reaction in the UV-Visible analysis indicating the complete reduction of methyl red^{26, 27}.

CONCLUSION

Biosynthesis gained huge fame in the scientific and research world for the synthesis of metallic nanoparticles because of its cost effective and ecofriendly nature. In the present study AgNPs were synthesized using the aqueous extract of *Passiflora foetida* aerial parts. The amalgamated solution of 10ml plant extract + 190ml AgNO₃ solution turned deep reddish brown in colour and has shown surface plasmon resonance band at 435.13nm in UV-Visible analysis after 48hrs confirming the formation of AgNPs. In DLS study the AgNPs have shown equal size distribution and no aggregation with mean size of 99.3nm. The plant mediated AgNPs have exhibited the zeta potential value of -20.8mV which proclaimed their moderately stable nature. The spectrum obtained in the XRD analysis clearly indicates that the biosynthesized AgNPs of *P. foetida* were crystalline face centred cubic in structure. The peaks obtained in the FTIR spectrum of AgNPs acknowledged the presence different photochemical in the plant extract of *P. foetida* and their role as reducing and stabilizing agents in AgNPs formation. The TEM images revealed that most of the AgNPs were spherical in shape with an average size of 20nm produced in XRD analysis. Further crucially in the present study the plant mediated AgNPs were examined for their catalytic activity in presence of NaBH₄ in the degradation and removal of 4-Nitrophenol, Methylene blue, Methyl orange and Methyl red. After adding AgNPs to the reduction reactions of 4-Nitrophenol, Methylene blue, Methyl orange and Methyl red the absorption maximum was decreased from 1.252 to 0.823, 2.009 to Nil, 1.673 to 0.647 and 2.651 to 0.281 respectively. The above results recorded in UV-Visible analysis confirmed that the AgNPs of *P. foetida* were shown to exhibit remarkable catalytic activity in the degradation and removal of respective synthetic and toxic chemicals.

CONFLICT OF INTEREST: All the authors declare that there is no conflict of interest

REFERENCES

1. Kjellstrom T, Madhumita L, Tony M. M, Geetha R, Rupendra S, Kingsland S. Air and Water Pollution: Burden and Strategies for Control. Disease control priorities in Developing countries 2006; 2(43): 817-832.

2. Muthu K, Priya S. Green synthesis, characterization and catalytic activity of silver nanoparticles using *Cassia auriculata* flower extract separated fraction. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy* 2017; 179: 66-72.
3. Rasheed T, Bilal M, Li C, Nabeel F, Khalid M, Iqbal H. M. N. Catalytic potential of bio-synthesized silver nanoparticles using *Convolvulus arvensis* extract for the degradation of environmental pollutants. *Journal of Photochemistry and Photobiology B: Biology* 2018; 181: 44-52.
4. Bodaiah B, Aswani Kumar Y. V. V, Vinay Viswanath K, Joy Richardson P, Usha Kiranmayi M, Sudhakar P. Anticancer activity of plant mediated silver nanoparticles on selected cancer cell lines, *Journal of Chemical and Pharmaceutical Research* 2016; 8(8):376-381.
5. Edison T. J. I, Sethuraman M. G. Biogenic robust synthesis of silver nanoparticles using *Punica granatum* peel and its application as a green catalyst for the reduction of an anthropogenic pollutant 4-nitrophenol. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy* 2013; 104: 262–264.
6. Bodaiah B, Usha Kiranmayi M, Aswani Kumar Y. V. V, Ravi Varma A, Vinay Viswanath K, Udaya Lakshmi K, Sudhakar P. Ecofriendly Synthesis of Silver Nanoparticles by using Indian Plants and Screening their Catalytic Activity. *Current Trends in Biotechnology and Pharmacy* 2016; 10(4): 316-323.
7. Bodaiah B, Usha Kiranmai M, Anuhya G, Yamini Saraswathi Y, Sambasiva Rao K. R. S, Sudhakar P. Green Synthesis of Silver Nanoparticles using two Apocyanaceae plants and Screening for their Catalytic Activity. *Current Trends in Biotechnology and Pharmacy* 2017; 11(1): 84-90.
8. Joseph A. P, Hemmalakshmi S, Priyanga S, Devaki K. Phytochemical screening, GC-MS analysis and enzyme inhibitory activity of *Passiflora foetida* L. *Indo American Journal of Pharmaceutical research* 2014; 4(8): 3526-3534.
9. Ravi Babu B, Jagadish Naik M, Siddappa Revana D, Jilani S. K, Janardhan M. Phytochemical screening of ethanolic extract of *Passiflora foetida*(Linn) and medicinal importance. *Indian Journal of Research in Pharmacy and Biotechnology* 2015; 3(4): 324-327.
10. Nalini M, Lena M, Sumathi P, Chandran S. Effect of phyto-synthesized silver nanoparticles on developmental stages of malaria vector, *Anopheles stephensi* and dengue vector, *Aedes aegypti*. *Egyptian Journal of Basic and Applied Sciences* 2017; 4: 212–218.
11. Vigneshwaran N, Ashtaputre N. M, Varadarajan P. V, Nachane R. P, Paralikar K. M, Balasubramanya R. H. Biological synthesis of silver nanoparticles using the fungus *Aspergillus flavus*. *Materials Letters* 2007; 61: 1413–1418.

12. Patil S, Chaudhari G, Paradeshi J, Mahajan R, Chaudhari B. L. Instant green synthesis of silver-based herbo-metallic colloidal nanosuspension in *Terminalia bellirica* fruit aqueous extract for catalytic and antibacterial applications. 3 Biotech 2017; 7:36.
13. Lim J. K, Yeap S. P, Che H. X, Low S. C. Characterization of magnetic nanoparticle by dynamic light scattering. Nanoscale Res Letters 2013; 1: 381.
14. Bodaiah B, Usha Kiranmayi, Vijayalakshmi M, Rama Krishna G. V. S, Sambasiva Rao K. R. S, Sudhakar P. Green synthesis, characterisation and biological activities of silver nanoparticles by using *Wrightia tomentosa* leaves. Journal of Chemical and Pharmaceutical Research 2016; 8(6):266-278.
15. Singh S, Bharti A, Meena V. K. Structural, thermal, zeta potential and electrical properties of disaccharide reduced silver nanoparticles. J. Mater. Sci .Mater. Elec. 2014; 25: 3747-3752.
16. Jyoti K, Baunthiyal M, Singh A. Characterization of silver nanoparticles synthesized using *Urtica dioica* Linn leaves and their synergistic effects with antibiotics. J of Radia. Research and Applied Science 2016; 9(3): 217-227.
17. Ashraf J. M, Ansari M. A, Khan H. M, Alzohairy M. A, Choi I. Green synthesis of silver nanoparticles and characterization of their inhibitory effects on AGEs formation using biophysical techniques. Scientific Reports 2016; 6:20414.
18. Suresh G, Gunasekar P. H, Kokila D, Prabhu D, Dinesh D, Ravichandran N, Ramesh B, Koodalingam A, Siva G. V. Green synthesis of silver nanoparticles using *Delphinium denudatum* root extract exhibits antibacterial and mosquito larvicidal activities. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 2014; 127: 61–66.
19. Ulug B, Turkdemir H. M, Cicek A, Mete A. Role of irradiation in the green synthesis of silver nanoparticles mediated by fig (*Ficus carica*) leaf extract. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 2015; 135 153–161.
20. Rahim K. A, Mahmoud S. Y, Ali A. M, Almaary K. S, Mustafa A. E. M. A, Husseiny S. M. Extracellular biosynthesis of silver nanoparticles using *Rhizopus stolonifer*. Saudi Journal of Biological Sciences 2017; 24: 208–216.
21. Li P, Li S, Wang Y, Zhang Y, Han G. Z. Green synthesis of β -CD-functionalized monodispersed silver nanoparticles with enhanced catalytic activity. Colloids and Surfaces A: Physicochemical and Engineering Aspects 2017; 520, 5: 26-31.
22. Shabir J, Rani S, Garkoti Ch, Mozumdar S. Nitrolotriacetic acid assisted one step synthesis of highly stable silver nanoparticles in aqueous medium: Investigation of catalytic activity. Materials Letters 2017; 209: 207-211.

23. Hamed S, Shojaosadati S. A, Mohammadi A. Evaluation of the catalytic, antibacterial and anti-biofilm activities of the *Convolvulus arvensis* extract functionalized silver nanoparticles. *Journal of Photochemistry and Photobiology B: Biology* 2017; 167: 36-44.
 24. Wang L, Lu F, Liu Y, Wu Y, Wu Z. Photocatalytic degradation of organic dyes and antimicrobial activity of silver nanoparticles fast synthesized by flavonoids fraction of *Psidium guajava* L leaves. *Journal of Molecular Liquids* 2018; 263: 187-192.
 25. Khan M. N, Bashir O, Khan T. A, AL-Thabaiti S. A, Khan Z. CTAB capped synthesis of bio-conjugated silver nanoparticles and their enhanced catalytic activities. *Journal of Molecular Liquids* 2018; 258: 133-141.
 26. Reddy N. J, Bhagat E, Suchiang K, Sadras S. R. Comparative Study of Antioxidant and Catalytic Activity of Silver and Gold Nanoparticles Synthesized from *Costus pictus* Leaf extract. *Journal of Materials Science and Technology* 2015; 31(10): 986-994.
 27. Umamaheswari C, Lakshmanan A, Nagarajan N. S. Green synthesis, characterization and catalytic degradation studies of gold nanoparticles against congo red and methyl orange. *Journal of Photochemistry and Photobiology B: Biology* 2018; 178: 33-39.
-