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### **Effect of Seasonal Variation and Edaphic Factors on am Population Associated With Medicinal Plants of Hardwar Range of Rajaji National Park, Uttarakhand**

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#### **ABSTRACT:**

“Mycorrhiza” – mark out a symbiotic association between a fungus and roots of the host plant, in which both are getting advantage. This association has importance in terms of growth, yield, nutrition, protection and biochemical contents of the host plant. These are omnipresent and fall under a broad ecological range. An investigation has been made to examine the spore density and root colonization of Arbuscular Mycorrhizal Fungi (AMF) in the rhizosphere of *Syzygium cumini* and *Atlantis monophylla* collected from Hardwar range of Rajaji National Park, Uttarakhand, India. The spore density ranges from 30-68/10gm and 46-345/10gm of soil whereas percent root colonization ranges from 30-70% and 20-80% for *Syzygium cumini* and *Atlantis monophylla* respectively. Maximum spore count and percent root colonization was recorded in rainy season and minimum was recorded in winter season. Six species were isolated, which belongs to genus *Glomus* and *Acaulospora*. *Glomus* was found to be predominant over *Acaulospora*. The edaphic factors were also analysed and correlated with the diversity measures.

**KEYWORDS:** Mycorrhiza, Symbiotic, Rhizosphere, *Glomus*, *Acaulospora*, Edaphic.

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## INTRODUCTION-

AM fungal spores multiply only in association with plant roots which act as a suitable ecological niche for germination of spores. Soil properties like texture, pH, moisture content, organic matter and phosphorus contents can be measured or determined and is used to characterise the soil inhabitant by a particular type of micro flora and fauna. Medicinal plants are a rich source of ingredients that are frequently used for drug synthesis. The World Health Organization (WHO) estimated that 80% of the population of developing countries relies on traditional medicines, mostly plant drugs, for their primary health care needs<sup>1</sup>. Because of the growing pressure of pharmaceutical industries, medicinal plants are being tilled. One fifth of all the plants in India are used for medicinal purpose. Majority of plants have the dynamic association with AM fungi. AM/VAM fungi occur in 80% of vascular plants<sup>2</sup>. AM fungi is a type of mycorrhiza, where the fungus resides in the cortical cells of the roots of the plant. They form unique structures viz. arbuscules and vesicles. It helps the plant to absorb nutrients from the soil. AM helps in the taking up of P and other nutrients from in sufficient area through fungal hyphae by expanding the root absorption area. AM fungi helps in soil conservation by making cluster of soil with the help of hyphal networks. AM hyphae can take up nutrients up to 12 cm from the surface of roots<sup>3</sup>. AM shows symbiotic relationship with the plants, which is the most common symbiosis. Symbiosis represents a mutual connection between two different living beings<sup>4</sup> in which both plant and fungus are complementary to each other. The conditions and growth of medicinal plants is improved by inoculation of AM in the root system. AM fungi associated with medicinal plants has not only enhanced the growth of these plants but also improved the active principle content<sup>5-6</sup>.

## EXPERIMENTAL SECTION

Most of the soil samples were collected from the rhizospheric soil of *Syzygium cumini* and *Atlantia monophylla* for the present study. Soil, a natural home for various microbes which maintains a dynamic equilibrium and its properties such as pH, phosphorus, organic carbon etc. has been determined as the distribution and occurrence of AM fungi vary with the change in edaphic factors. The objective of this study is to find out the species diversity of AM fungi in rhizospheric soil with different physiochemical properties.

**Study site-** Studies on arbuscular mycorrhizal fungi of rhizospheric soil were carried out on medicinal plants, collected from Hardwar range of Rajaji National Park, Uttarakhand India. It is located at an altitude of 302-1000m asl and falls under sub tropical moist deciduous forest type.

**Soil sample collection-** Soil and root samples were collected from the site using soil auger at a depth of 5-15cm. It is stored at room temperature for further analysis.

**Study of root colonization** – Roots were washed and cut into 1cm pieces in length for determination of percent mycorrhizal colonization<sup>7</sup>.

$$\% \text{ root colonization} = \frac{\text{Total number of roots segments colonized}}{\text{Total number of roots segments examined}} \times 100$$

**Isolation and identification of AM fungi**- 10 gm of rhizospheric soil samples were analysed for spore isolation by wet sieving and decanting technique<sup>8</sup>. The isolated AM spores were identified on morphological basis with help of monograph given by Schenck and Perez<sup>9</sup> and INVAM (<http://www.invam.caf.wvu.edu>).

**Physiochemical analysis of soil samples**- As the AM fungi was influenced by soil parameters, physiochemical analysis of soil samples were also done.

**Soil pH**- 1 gm of soil is dissolved in 10ml of DW to make a suspension. Further, the supernatant was taken and the pH of the soil was measured using pH meter.

**Chemical properties**- Soil organic carbon<sup>10</sup>; Phosphorus<sup>11</sup> and Potassium<sup>12</sup> were determined.

**Statistical analysis** –Spore density and root colonization were measured. Pearson's correlation coefficient was used to assess the relationship between diversity measures and edaphic factors.

## RESULT AND DISCUSSION-

During the present investigation, AM fungal spores were found to be well distributed in all soil samples analysed as well as roots studied for root colonization although their types and number varied considerably. Both medicinal plants showed varied number of spores (Table 1).

**Table 1. Soil parameters in test plants**

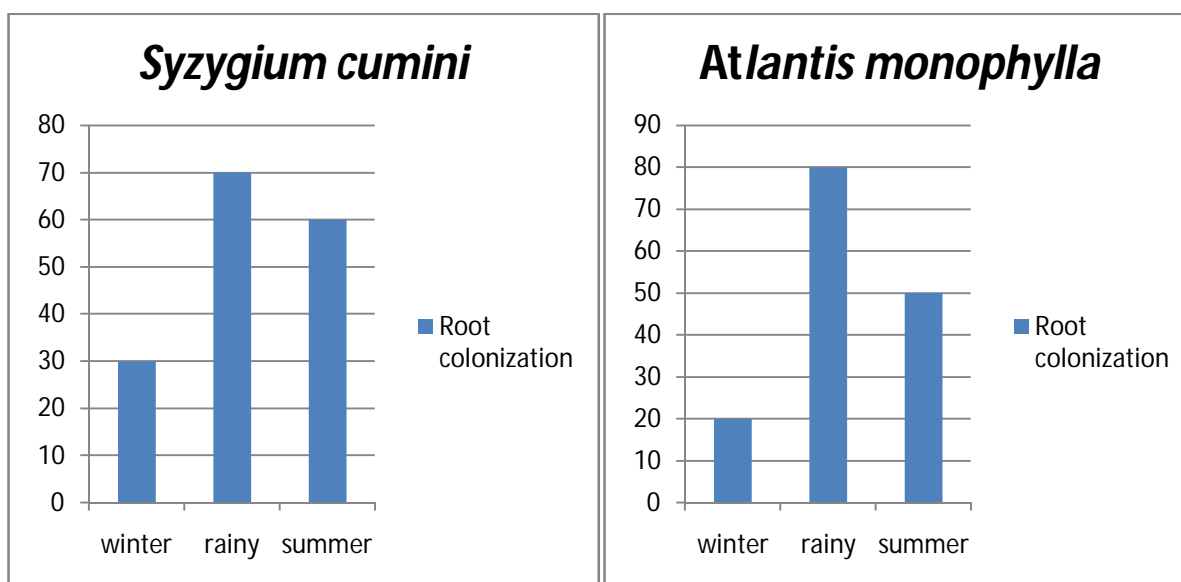
Plant species	Season	Spore count	% Root colonization	pH	O.C	P	K
<i>Syzygium cumini</i>	Winter	30±7.72	30%	6.9±0.26	0.06±0.02	14±2.51	119±3
	Rainy	68±7.54	70%	7.4±0.30	0.57±0.03	8.7±2.51	155±2.04
	Summer	42±7.63	60%	6.9±0.25	0.70±0.02	13.5±2.6	310±3
<i>Atlantis monophylla</i>	Winter	46±5.56	20%	6.9±0.26	0.09±0.02	9±2.51	121±3.05
	Rainy	345±10.96	80%	7.2±0.25	0.64±0.03	13.8±2.31	178.9±3.63
	summer	147±4.50	50%	7±0.3	0.51±0.02	19.7±2.08	192.9±4.11

During present study, spore population ranges from 30-68 in *Syzygium* and 46-345 in *Atlantia*. Percent root colonization ranged from 30-70% in *Syzygium* and 20-80% in *Atlantia* respectively (Fig.1 a, b). Rhizospheric soil samples were processed and number of AM spores/10 gm of soil were assessed which range from 30-345. *Atlantia* showed high spore count as compared to *Syzygium* in all seasons. 6 species belonging to two genera namely *Glomus* and *Acaluospora* were observed (Fig. 3). *Glomus* was found to be the most dominant species but *Acaluospora* species abundance were also evident. Table 1 depicts soil pH, organic carbon, phosphorus and potassium

showing effect on spore population and root colonization. In case of *Syzygium*, it is evident that the soil pH ranges from 6.9-7.4 and shares a strong positive relation with spore count and percent root colonization. The O.C content ranges from 0.06-0.70 and shares a strong positive correlation with both spore count and root colonization. Phosphorus and potassium shares a strong negative correlation with both measures whereas *Atlantia* study revealed its pH range from 6.-7.2. O.C content was negatively correlated with spore count and positively correlated with root colonization. Potassium and phosphorus shares a positive correlation with spore count and a negative correlation with root colonization. Correlation values of soil properties with respect to spore count and root are calculated. The spore density and root colonization of both plants are highly correlated (Table 2).

**Table 2. Correlation between different parameters of test plants**

	pH	O.C	P	K
<i>Syzygium cumini</i> -spore count	0.951101277	0.6006086	-0.97401213	-0.6312796
<i>Atlantis monophylla</i> -spore count	-0.9829076	-0.042922105	0.725875048	0.34746562
<i>Syzygium cumini</i> percent colonization	0.69337524	0.90648412	-0.752400317	-0.443351699
<i>Atlantis monophylla</i> percent colonization	0.65465367	0.226118305	-0.586839577	-0.16889784



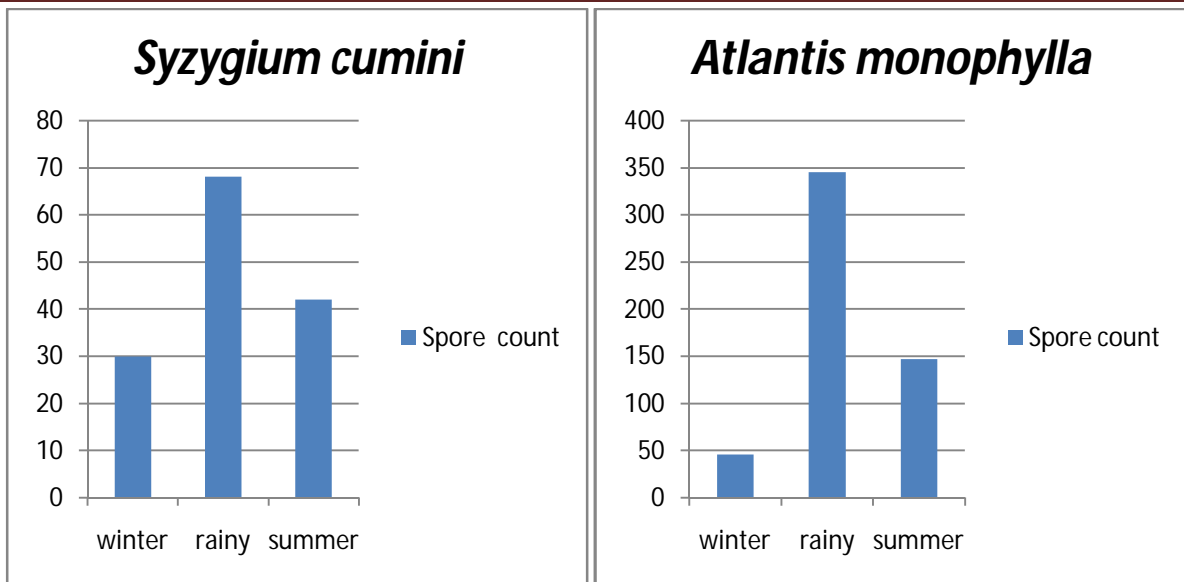


Fig. 1 Seasonal variation in test plants

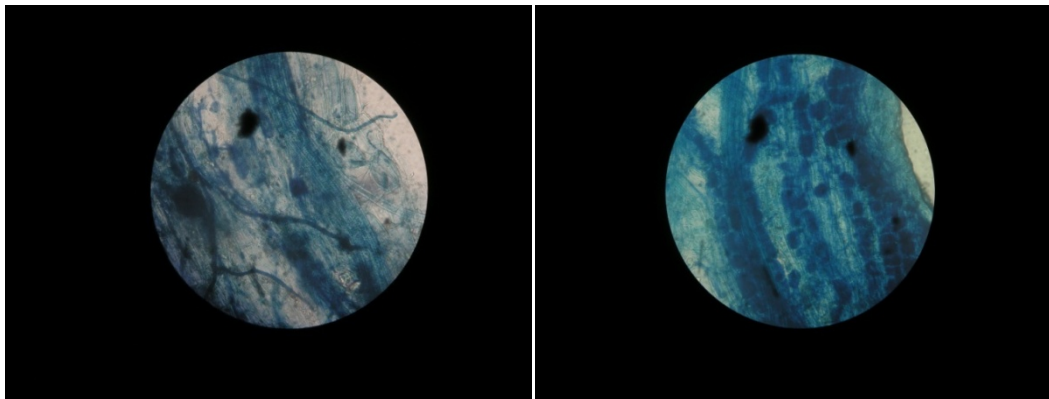
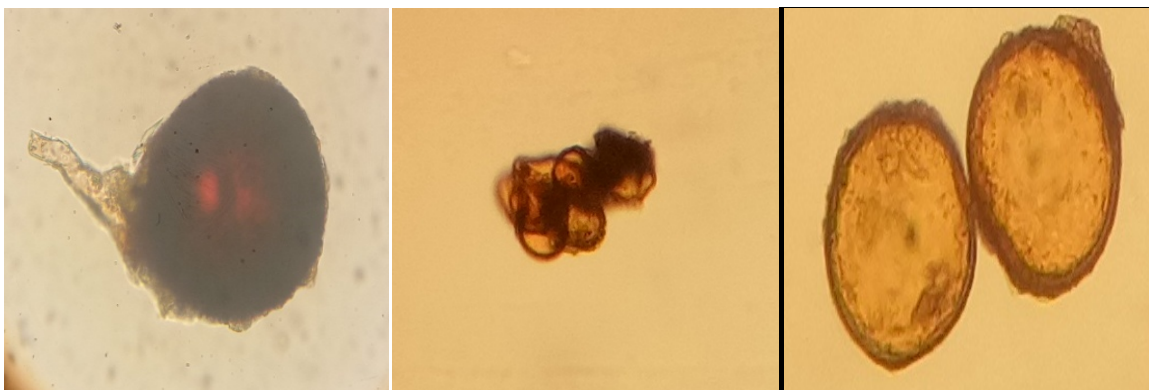


Fig. 2 Root colonization by AMF spores



*Glomus fasciculatum*

*Glomus fasciculatum*

*Acaluospora mella*

Fig. 3- Isolated spores from rhizospheric soil

Soil is a constantly changing medium activated by the interplay of a vast variety of macroscopic and microscopic forms of life. AM fungi have a widespread distribution throughout the plant kingdom and the association of fungi is geographically ubiquitous. The extent of root

colonization and spore number is the criteria for determining AM diversity and richness. AM has directly effect on structure and diversity of plant species. In the present study, spore population ranges from 30-68 in *Syzygium* and 46-345 in *Atlantia*. Also, the percent root colonization range from 30-70 in *Syzygium* and 20-80 in *Atlantia*. *Glomus* was the predominant species followed by *Acaluospora*. Both plant shows high spore density in rainy season followed by summer and winter. Root colonization also shows the same pattern -high in rainy season, moderate in summer season and lowest in winter. These results are in agreements with the earlier reports<sup>13-17</sup>. The predominant species is *Glomus* which is related with earlier observations<sup>18-21</sup>. Soil factors directly affect the mycorrhizal population. pH shares a positive correlation with *Syzygium* and negative with *Atlantia* which are related to Wang et al.<sup>22-23</sup>. AM spores increase with increasing organic carbon which are in accordance with Johnston<sup>24</sup>. Phosphorus is negatively correlated with both plants which are in agreements with Javadi et al.<sup>25</sup>. Potassium also shows negative correlation which are related to Khanam et al.<sup>26</sup>.

## CONCLUSION

AM fungi are present in soil in the form of both sexual and asexual spores i.e. zygospores, azygospores and chlamyospores and also in the form of hyphae and vesicles associated with plant roots. Each of these forms remains dormant in soil till they find a suitable host for growth and reproduction. During the present study, the AM fungi revealed very extensive role for understanding the dynamics of these fungi in soil. As per the result of present study, it can be concluded that both seasons and soil parameters affects the AM spore density. Rainy season is given preference as compare to winter and summer in case of spore count and root colonization but we can't neglect the environmental factors like pH, O.C etc.

## REFERENCES

1. Canter PH, Thomas H and Ernst E. Bringing medicinal plants into cultivation: opportunities and challenges for biotechnology. Trends in Biotech.2005; 23:180-185.
2. Harley JL and Harley EL. A check list of Mycorrhiza in the British Flora. New Phytol.1987; 105: 1-102.
3. Cui M and Caldwell MM. Facilitation of plant phosphate acquisition by arbuscular mycorrhizas from enriched soil patches II. Hyphae exploiting root-free soil. New Phytol.1996; 133(3):461-467.
4. Gaur S and Kaushik P. Biodiversity of vesicular arbuscular mycorrhiza associated with *Catharanthus roseus*, *Ocimum* spp. and *Asparagus racemosus* in Uttarakhand State of Indian Central Himalaya. Int. J. Bot. 2011a; 7(1): 31-41.



5. Basu M and Srivastava NK. Root endophytes in medicinal plants: Their population and effect ICCP 98 G.T.S. Baylis, 1967. Experiments on the ecological significance of phycomycetous mycorrhizas. *New Phytol.*1998; 66:231-243.
6. Zubek S and Blaszkowski J. Medicinal plants as hosts of arbuscular mycorrhizal fungi and dark septate endophytes. *Phytochem. Rev.* 2009; 8:571-580.
7. Philips JM and Hayman DS. Improved procedures for clearing roots and staining parasitic and vesicular arbuscular mycorrhizal fungi for rapid assessment of infection. *Trans. British Mycol. Soc.*1970; 55(1):158-161.
8. Gerdemann JW and Nicolson TH. Spores of mycorrhizal Endogone species extracted from soil by wet sieving and decanting. *Trans. British Mycol. Soc.*1963; 46: 235-244.
9. Schenck NC and Perez Y. Manual for the identification of VA mycorrhizal fungi. Synergistic Publications, University of Florida, Gainesville, Florida, U.S.A. 1989.
10. Walkley A and Black JA. An examination of the Degtjareff method for determining soil organic matter and a proposed modification of the chromic titration method. *Soil Sci.*1934;37:29–38.
11. Olesen SR, Cole CV, Watanable FS and Dean LA. Estimation of available phosphorus in soil by extraction with sodium bicarbonate. USDA circular no. 939. US govt. Printing office, Washington , DC 1954;1-19.
12. Hanway JJ and Heidel H. Soil analysis method as used in Iowa state college soil testing laboratory, Iowa State College of Agriculture;1952:1-31.
13. Bhaskaran C and Selvaraj T. Season incidence and distribution of VA-Mycorrhizal fungi in native saline soils. *J. Env. Biol.*1997; 18:209-212.
14. Kumar R, Tapwal A, Jaime A et al. Observations on arbuscular mycorrhiza associated with important edible tuberous plants grown in wet evergreen forest in Assam, India. *Biodiversitas.* 2013; 14(2): 67-72.
15. Khamar Jahan MD, Bavaji M and Sreeramula A. Occurrence of AM fungi in rhizosphere soil of endemic and endangered medicinal plants. *Ind. J. Fund. Appl. Life Sci.* 2012; 2(2): 276-280.
16. Chanda D, Sharma GD and Jha DK. Isolation and identification of some Arbuscular Mycorrhiza (AM) fungi for phytoremediation in soil contaminated with paper mill effluent, Silchar, Assam, India. *Int. J. Curr. Microbiol. Appl. Sci.*2014; 3(6):527-539.
17. Jaya T and Shinde BP. Studies on rhizobium and AM fungi associated with Pea (*Pisum sativum* L.). Ph.D. Thesis, University of Pune.2015.

18. Lakshman HC and Ratna V. Seasonal fluctuation of AM fungi on two hydrocarbon yielding plant species of *Jatropha*. *Int. J. Adv. Res.* 2015; 3(7):379-385.
19. Mosse B. Advances in the study of vesicular-arbuscular mycorrhiza. *Ann. Rev. Phytopath.* 1973; 11:171-196.
20. Khade SW and Rodrigues BF. Occurrence of arbuscular mycorrhizal fungi in tree species from Western Ghats of Goa, India. *J. Trop. For. Sci.* 2003a; 15:320-331.
21. Bukhari MJ, Khade SW, Jaiswal V, Gaonkar UC and Rodrigues BF. Arbuscular mycorrhizal status of medicinal plants: A field survey of AM fungal association in herbs. *Plant Archives.* 2003; 3:167-174.
22. Wang GM, Stribley DP and Tinker PB. Soil pH and vesicular arbuscular mycorrhizas in ecological interactions in soil. Fitter A H (ed.) Oxford Blackwell Scientific Publications. 1985; 219-224.
23. Wang GM, Stribley DP, Tinker PB and Walker C. Effects of pH on arbuscular mycorrhiza field observations on the long term liming experiments at Roathamsted and Woburn. *New Phytol.* 1993; 124:465-472.
24. Johnston A. Vesicular arbuscular mycorrhiza in sea island cotton and other tropical plants. *Trop. Agri. Trinidad.* 1949; 26:118-121.
25. Javadi M, Beuerleln JE and Arscott TG. Effects of phosphorus and copper on factors influencing nutrient uptake, photosynthesis and grain yield of wheat. *Ohio J of Sci.* 1991; 91:191-194
26. Khanam D, Mridha MAU, Soliaman ARM and Hossain T. Effects of edhaphic factors on root colonization and spore population of arbuscular mycorrhizal fungi. *Bull. Inst. Trop. Agric. Kyushu. Univ.* 2006; 29:97-104.