

International Journal of Scientific Research and Reviews

Extraction of Pigment from *Rhodotorula Mucilaginosa* and its Application in Textile

Hemashenpagam.N^{1*}, Karuppusamy.R² and Pratheeba .M³

^{1*} Associate Professor, PG and Research Department of Microbiology

^{2,3} II M. Sc Microbiology, Hindusthan college of Arts and Science Coimbatore-641028

ABSTRACT

Yeasts are more convenient than algae or molds for large scale production in fermenters, due to their unicellular nature and high growth rate. The present investigation was undertaken with an aim to identify and characterize pigment from yeast. Yeast were isolated from kitchen waste mixed soil sample collected from Avinashi, Tirupur, Tamil Nadu, India. The potent strain (KAR1) *Rhodotorula mucilaginosa*. The extraction of pigment was done by solvent extraction method using acetone and purified by column chromatography. Mango bark used as a mordant. Antibacterial activity of pigment, mordant and pigment with mordant inhibitory action against *E. coli*, *Pseudomonas*, *Staphylococcus aureus*, *Klebsiella pneumonia*, *Proteus*, *Bacillus cereus* were checked. Antifungal activity against fungal pathogens like *Candida albicans* and *Trichoderma* were tested against microbial pigment, mordant and dye + mordant to evaluate its antifungal activity. As a source of natural pigment these pigments were used for textile dyeing.

KEYWORDS: Natural pigment, microbial pigment, yeast, extraction of pigment, antimicrobial activity, Textile application

***Corresponding author:**

Hemashenpagam. N

Associate Professor,

PG and Research Department of Microbiology

Coimbatore-641028

INTRODUCTION

The toxicity problems caused by synthetic dyes to the environment have created amounting interest towards natural pigments. Pigments from microbial sources as natural dyes are potentially good alternative ones to synthetic pigments. There are several organisms which produce many varieties of intracellular and extracellular pigments including melanin with different biological functions. Pigments were primarily used as a colouring agent in various industries from the past decade. Researchers have focused the usage of natural pigments for various industries including pharmaceutical for antitoxic and antioxidant agents¹

Most of the natural pigments are extracted from plants like annatto, beet root, marigold, grapes, carrot, paprika, etc. and microorganisms like yeast of the genera *Phaffia*, *Cryptococcus* and *Sporobolomyces*, fungi like *Blakesleatrispora*, *Monascus sp.*, and algae such as *Dunaliella* and *Haematococcus* and bacteria such as *Flavobacterium* and *Micrococcus* are reported to produce carotenoids² Pigmented yeasts like *Rhodotorula* and *Rhodospiridium* produce the major carotenoid pigments viz. carotene, torulene, torularhodin.

Yeasts can synthesize pigment when cultivated in commercial medium, containing various carbon sources, such as glucose, xylose, cellobiose, sucrose, glycerol and sorbitol nevertheless these type of medium represents high costs. Therefore there have been growing interest in the use of natural substrates as carbon sources³ By-products from industrial processes are pollutants to the environment and their treatment represents high costs. In recent years raw materials and agro-industrial wastes origin have been proposed as low-cost alternative carbohydrate sources.

The textile industry discharges large proportion of effluent that mainly consists of synthetic dyes. Synthetic dyes been extensively used in the textile industries due to their ease and cost effectiveness, high stability towards light, temperature and technically advanced colours covering the whole colour spectrum. However, these synthetic dyes are often toxic, mutagenic and carcinogenic leading to several human health problems such as skin cancer and allergic reactions^{4,5}. Thus, the worldwide demand for the dyes of natural origin is increasing rapidly in the textile industry.

MATERIALS AND METHODS

Sample collection:

Soil samples were collected from different areas of Avinashi, Tirupur, Tamil Nadu, India. Kitchen waste mixed with soil and fruits and vegetable waste composted soil were used

Isolation of yeast cells:

Isolation of yeast cells was performed by serial dilution and spread Plate technique using Yeast malt agar(YMA) and Potato dextrose agar(PDA). Plates were incubated at 30°C temperatures at 3 to 4 days.

Screening of pigment producing yeasts:

Yeast strains were isolated in this study and screen the efficient isolates. Potential isolates were chosen from primary screening and subjected through staining technique (Lacto phenol cotton blue stain). Morphological characteristics were detected.

Fermentation process of pigment producing yeasts:

The pigmented yeast isolate was incubated into production media YMB(Yeast malt broth) at 30°C for 7 Days on a rotatory shaking incubator at 100 rpm. YMB –Yeast Malt Broth: Dextrose-10g, Peptone-5g, Malt extract-3g, Yeast extract-3g, Distilled water-1000ml.

Extraction of pigment:

The yeast culture was inoculated on fermentation broth and incubated at 25 °C for 5 days. A known amount (500mg) of freeze-dried red yeast was hydrolyzed with 1 ml of 1N hydrochloric acid in water bath at 70 °C for one and half hour. After removal of excess acid by washing with water, the cells were soaked overnight in acetone: methanol (1:1) solution. The pigment was extracted with acetone until the entire colour was leached out from the cells. Acetone extracts were transferred to light petroleum (20ml) at (40-60 °C) in a separating funnel and washed thrice with distilled water. The pigment was collected and stored at 4°C.

Purification of pigment:

Using a silica gel chromatography column, the extracted solution was separated, and the pigments were eluted using hexane. It consists of glass tube with bottom portion of the column packed with glass wool/cotton. Above which absorbent is packed. Stationary phase (absorbent) – Silica gel was packed in the column. Sand was loaded in the top of the cotton and then silica gel was then packed the column. The crude extract was loaded at the top of the column and eluted using ethyl acetate as solution system. Fraction was collected at 20 minutes intervals. The fraction is further check qualitative and quantitative analysis.

Mordant preparation:

5gm of Mango bark powder is dissolved in 100ml of distilled water and boiled for 1hr. Then the extract was filtered using Whatmann no. 1 filter paper.

Antimicrobial activity:

Antibacterial activity of the pigment, mordant and pigment + mordant was tested by well diffusion method. Some pathogenic bacterial like (*E. coli*, *Pseudomonas*, *Staphylococcus aureus*, *Klebsiella pneumonia*, *Proteus*, *Bacillus cereus*.) were used against extracted Pigment, mordant and dye + mordant to evaluate its antibacterial activity. Then the wells were filled with appropriate amount of samples (50 µl) and it was incubated at 37°C for 24 hours and the result was observed by measuring zone of inhibition.

Antifungal activity of the pigment, mordant and pigment + mordant was tested by well diffusion method. Some fungal pathogenic like *Candida albicans* and *Trichoderma* were used against microbial pigment, mordant and dye + mordant to evaluate its antifungal activity. Then the wells were filled with appropriate amount of samples (50 µl) and it was incubated at room temperature for 3 days and the result was observed by measuring zone of inhibition.

Dyeing experiment:

A textile material (cotton) which is commercially available was selected for the experiment. Material was cut into equal size of 5 cm. Pigment in acetone was used as the stock solution. From this stock solution 5 ml solution was applied to the cloth material in a warm surface. The cloth material was allowed to dry at room temperature for about 1 hour. A white cloth material was taken as a control.

Washing performance:

The textile material dyed by pigment was tested for wash performance at room temperature. The textile material was washed with soap solution for 30 minutes at room temperature. The textile materials were washed with running tap water and allowed drying. The result was observed physically with other dyed unwashed textile material⁶.

Molecular identification of yeast:

Genomic organization of the 18srRNA and ITS1 sequencing

RESULT AND DISCUSSION

Sample collection and isolation of Pigmented yeast:

Soil samples were collected from different areas of Avinashi, Tirupur, Tamil Nadu, India. Kitchen waste mixed with soil samples and fruits and vegetable waste composted soil samples were collected and processed. The Samples were processed in YMA, PDA and SDA media's. Six different yeast strains were identified. The potent strain was found to be pigmented producing yeast(*Rhodotorulamucilaginos*). (Shows figure 1).

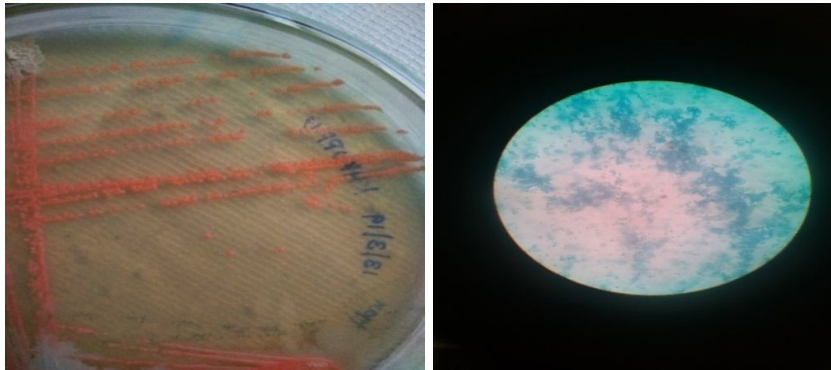


Figure 1 *Rhodotorulamucilaginos* in plate YMA and LCB Staining

Fermentation Process for Pigment Production

The isolated pigmented yeast was incubated in production media consists of Yeast Extract - 3g, Malt Extract -3g, Dextrose -10g, Peptone -5 g, Distilled Water -1000ml. (pH 6) at 30°C for 4 to 8 Days on a rotatory shaking incubator at 100 rpm for fermentation process.

Extraction of pigment:

The yeast culture was inoculated on fermentation broth and incubated at 8 to 10 days. A known amount (500mg) of freeze-dried red yeast was hydrolyzed with 1 ml of 1N hydrochloric acid in water bath at 70 °C for one and half hour was shown in figure 2. After removal of excess acid by washing with water, the cells were soaked overnight in acetone: methanol (1:1) solution. The pigment was extracted with acetone until the entire colour was leached out from the cells was shown in the figure 3. Acetone extracts were transferred to light petroleum (20ml) at (40-60 °C) in a separating funnel and washed thrice with distilled water. The pigment was collected and stored at 4°C. (Figure 4 shows the extracted pigment).

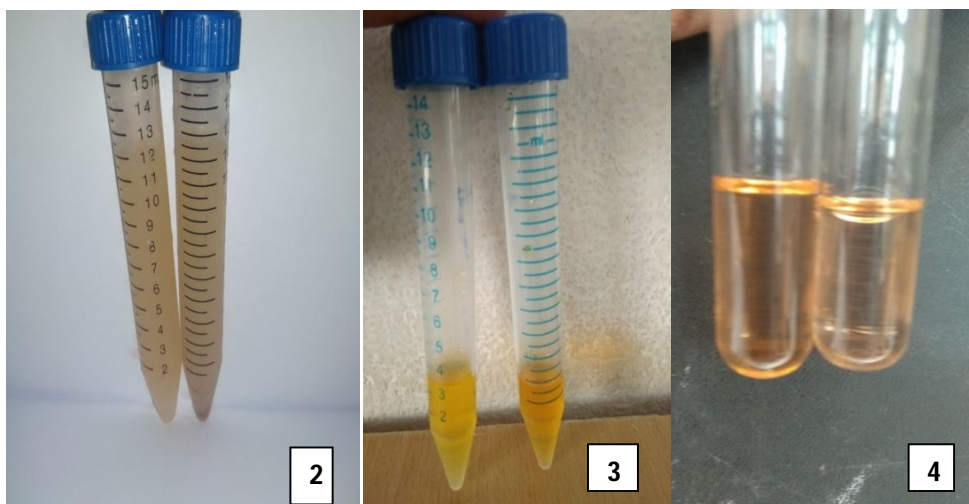


Figure 2, 3 and 4 Pigment extraction process

Purification of Pigment

Using a silica gel chromatography column, the extracted solution was separated, and the pigments were eluted using hexane. It consists of glass tube with bottom portion of the column-packed with glass wool/cotton. Above which absorbent is packed. Stationary phase (absorbent) – Silica gel was packed in the column. Sand was loaded in the top of the cotton and then silica gel was then packed the column was shown in the figure 5.4. The crude extract was loaded at the top of the column and eluted using ethyl acetate as solution system. (Figure 5 shows the purification dye). Fraction was collected at 20 minutes intervals were shown in figure 5. The fraction is further check qualitative and quantitative analysis.

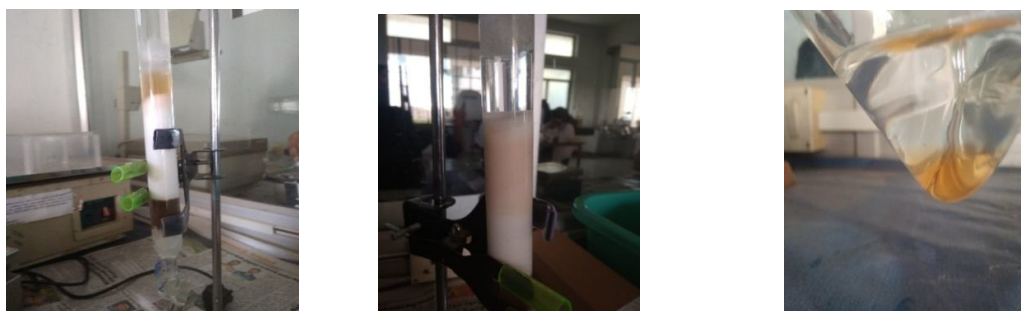


Figure 5 Column chromatography

Mordant Preparation

5gm of Mango bark powder is dissolved in 100ml of distilled water and boiled for 1hr. Then the extract was filtered using Whatman no. 1 filter paper.

Antimicrobial Activity

Antibacterial activity of the pigment, mordant and pigment + mordant was tested by well diffusion method. Some pathogenic like (*E. coli*, *Pseudomonas*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Proteus*, *Bacillus cereus*.) were used against extracted Pigment, mordant and dye + mordant to evaluate its antibacterial activity. Then the wells were filled with appropriate number of samples (50 µl) and it was incubated at 37°C for 24 hours and the result was observed by measuring zone of inhibition in Figure 6

Klebsiella pneumonia is found to be more resistant for microbial pigment than compare to *E. coli*, *Pseudomonas*, *Staphylococcus aureus*, *Proteus* and *Bacillus cereus*. *E. coli* is found to be more resistant for mordant than compare to *Pseudomonas*, *Staphylococcus aureus*, *Klebsiella pneumonia*, *Proteus*, *Bacillus cereus* in Figure 6..

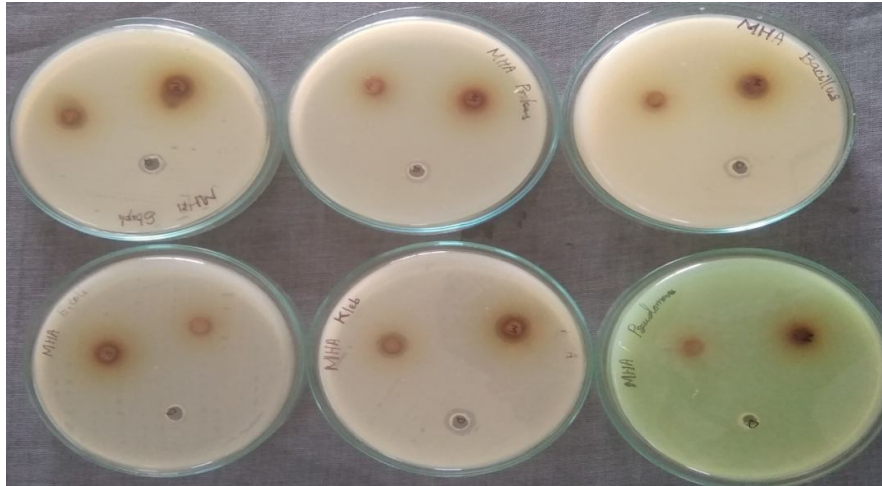


Figure 6 Antibacterial activity of Dye, mordant and Dye+mordant

Table 1 Antimicrobial activity of pigment extract with and without mordant

ORGANISMS	PIGMENT(DYE)	MORDANT	DYE+MORDANT
<i>S. aureus</i>	8mm	12mm	13mm
<i>Klebsiella pneumonia</i>	10mm	13mm	11mm
<i>Proteus sp.,</i>	6mm	12mm	8mm
<i>E.coli</i>	8mm	25mm	18mm
<i>Bacillus sp.,</i>	8mm	11mm	11mm
<i>Pseudomonas sp.,</i>	4mm	11mm	9mm

Antifungal Activity

Antifungal activity of the pigment, mordant and pigment + mordant was tested by well diffusion method. Some fungal pathogenic like *Candida albicans* and *Trichoderma* were used against microbial pigment, mordant and dye + mordant to evaluate its antifungal activity. Then the wells were filled with appropriate amount of samples (50 µl) and it was incubated at room temperature for 3 days and the result was observed by measuring zone of inhibition. (The Antifungal activity of dye, Mordant, Dye + Mordant was shown in the figure 7).

Candida albicans is found to be more sensitive for microbial pigment, microbial pigment + mordant and mordant than compare to *Trichoderma*. (figure shows 7).

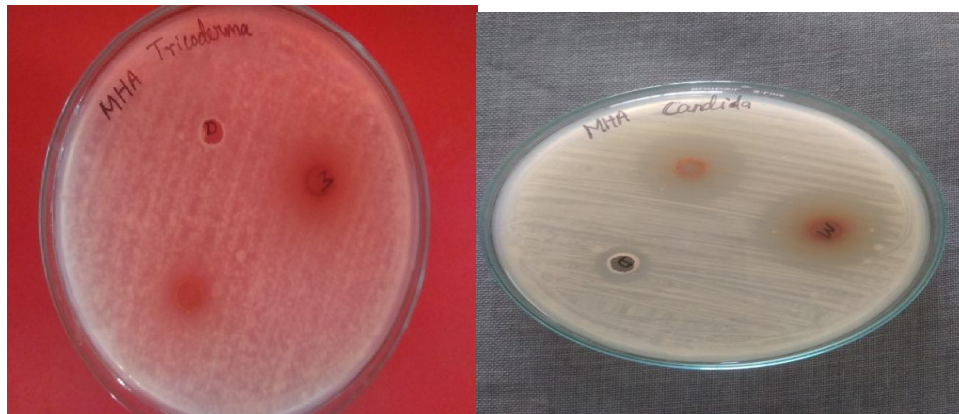


Figure 7 Antifungal Activity of Dye, Mordant and Dye+Mordant

Table 2: Antifungal activity in dye with and without mordant

ORGANISMS	PIGMENT(DYE)	MORDANT	DYE+MORDANT
<i>Candida</i>	12mm	25mm	20mm
<i>Trichoderma</i>	0mm	17mm	11mm

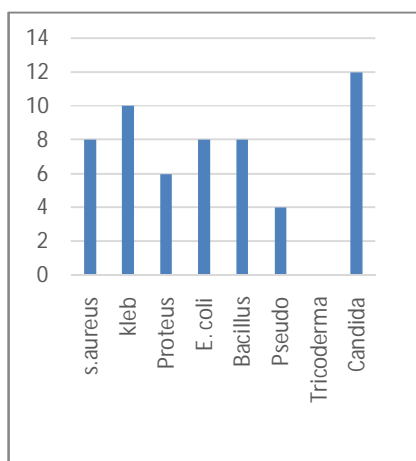


Figure 8 Zone of inhibition in microbial Pigment

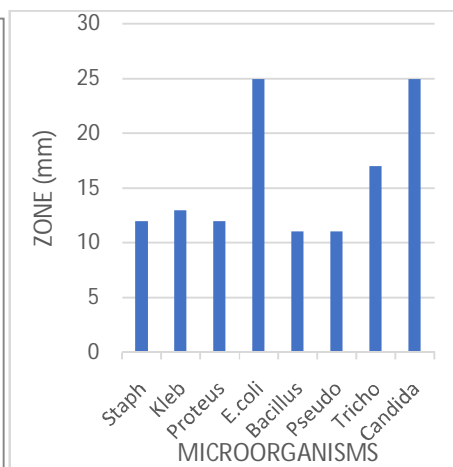


Figure 9 Zone of inhibition in Mordant

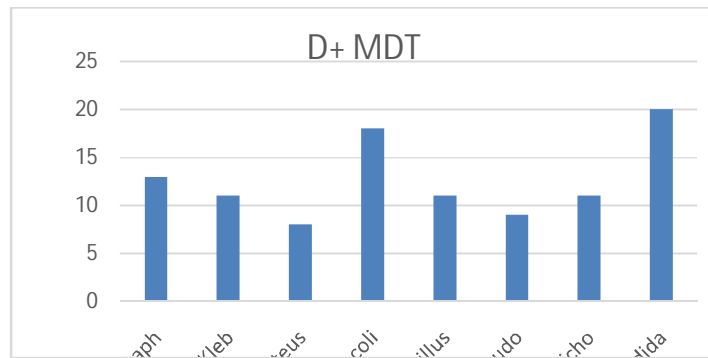


Figure 10 Zone of inhibition of dye and mordant

Figure 8,9,10 Antimicrobial activity of Microbial dye, Mordant and Microbial dye+ Mordant against test organisms

Dyeing Experiment

A textile material (cotton) which is commercially available was selected for the experiment. Material was cut into equal size of 5 cm. Pigment in acetone was used as the stock solution. From this stock solution 5 ml solution was applied to the cloth material in a warm surface. The cloth material was allowed to dry at room temperature for about 1 hour. A white cloth material was taken as a control. Figure 11 shows Pigmented cloth materials before washing.

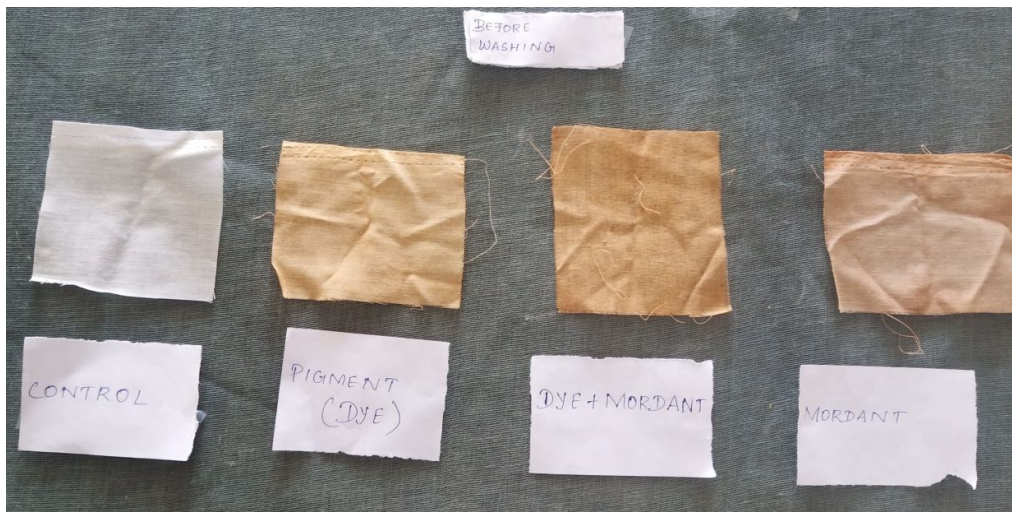


Figure 11 Pigmented cloth material before washing

Washing performance:

The textile material dyed by pigment was tested for wash performance at room temperature. The textile material was washed with soap solution for 30 minutes at room temperature. The textile material was washed with running tap water and allowed to dry. The result was observed physically with other dyed unwashed textile material.

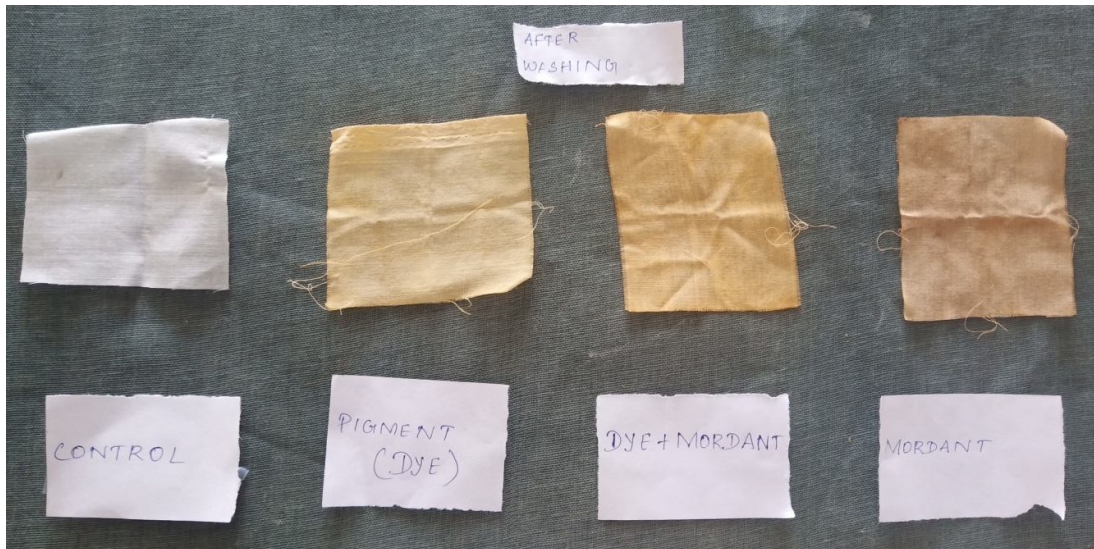


Figure 12 Pigmented cloth material after washing

Dye + Mordant was found to be effective then mordant and dye alone. It retains the orange colour after washing with soap solution. Then (Dye) orange colour decolorized to sandal colour was found to be retentive for the textile materials shown in figure 12.

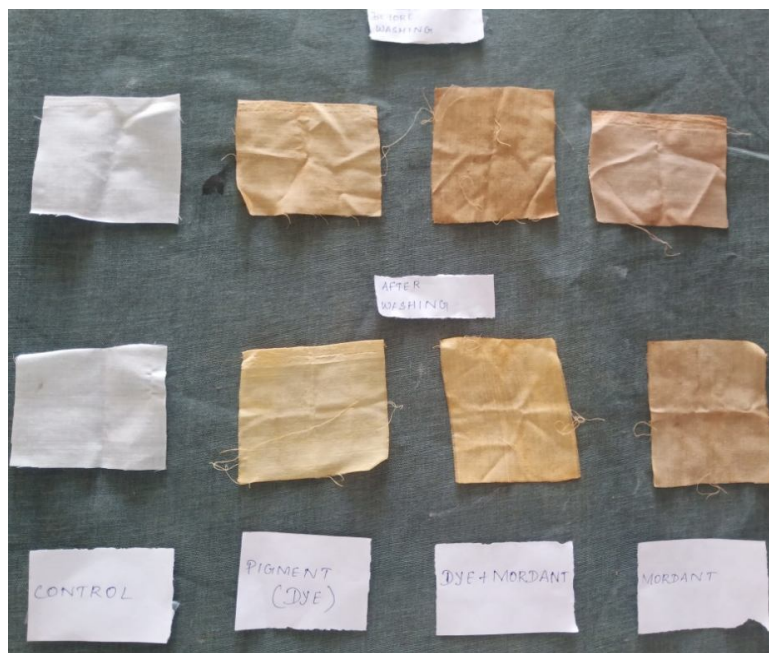


Figure 13 Comparison (Before Washing and After Washing) Of Microbial Dye In Cloth And After Washing

The textile material was used to check the dyeing property of the extracted pigment. The textile material after dyeing was subjected to three consecutive normal water wash treatments and detergent water wash treatment. Then orange colour decolorized to sandal colour was found to be retentive for the textile materials. (Figure 13). In textile industries, these pigments extracted from biological

source can be used as an alternative to the synthetic colorants and also which are safe and cost effective^{7,8}.

Molecular Identification of Yeast

Genomic organization of the 18srRNA and ITS1 sequencing were performed. DNA is isolated and further subjected to PCR Amplified. Then read the primers and then submitted in BLAST. The organism is confirmed by above using sequencing method as *Rhodotorulamucilaginos*⁹(Figure 14)

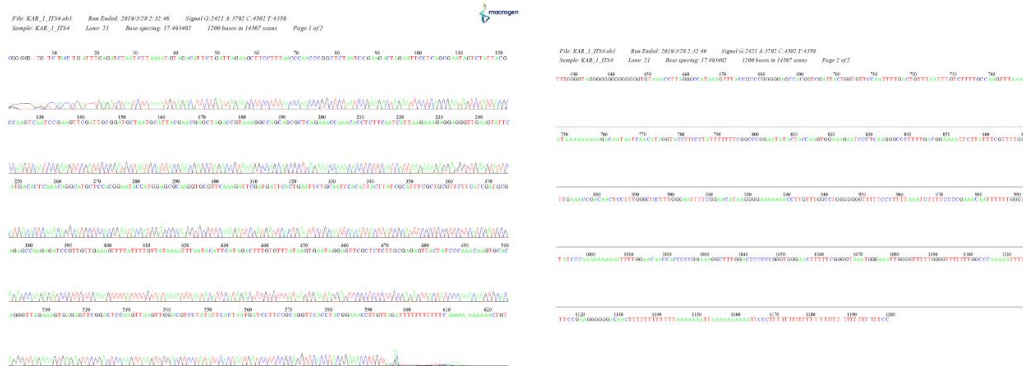


Figure 14 Molecular Identification of yeast

BLASTN 2.6.0+

Database: Nucleotide collection (nt)

43,634,103 sequences; 150,987,822,336 total letters

Query= 190327-R04_I05_KAR_1_ITS1 1 1200

Length=1200

	Score	E
Sequences producing significant alignments:	(Bits)	Value
KC113304.1 Rhodotorulamucilaginos	1044	0.0
LT548978.1 Rhodotorulamucilaginos	1042	0.0
LT548977.1 Rhodotorulamucilaginos	1042	0.0
KF726105.1 Rhodotorulamucilaginos	1042	0.0
KC113312.1 Fungal sp. PKU Y9 18S	1042	0.0
KR912272.1 Rhodotorulamucilaginos	1040	0.0
KT876700.1 Rhodotorulamucilaginos	1040	0.0
KC113310.1 Rhodotorulamucilaginos	1040	0.0
LT548979.1 Rhodotorulamucilaginos	1038	0.0
LC229714.1 Rhodotorula sp. EY12114	1038	0.0

- [KY104794.1](#)Rhodotorulamucilaginosa culture-collection CBS:10946... [1038](#) 0.0
[KT876501.1](#)Rhodotorulamucilaginosa isolate X5-3 internal transc... [1038](#) 0.0
[KF411538.1](#)Rhodotorulamucilaginosa strain DY115-21-1-Y46 intern... [1038](#) 0.0
[KF411535.1](#)Rhodotorulamucilaginosa strain DY115-21-1-Y38 intern... [1038](#) 0.0
[DQ186608.1](#) Rhodotorula sp. Y11 18S ribosomal RNA gene, partial s... [1038](#) 0.0
[LC277143.1](#)Rhodotorulamucilaginosa genes for SSU rRNA, ITS1, 5.... [1037](#) 0.0

CONCLUSION

In the near future, the product with natural colors may have an increased demand, not only for the safety of health and environment but also for their beauty and novelty. Increased awareness for eco-friendly products in the developed countries has opened up a new channel for the export of hand printed fabrics printed with natural dyes. Natural colors should not be taken as a threat to synthetic colors. It may take decades to manufacture natural colors in a ready to use form if all it is possible. A very long and consistent effort is required, since we have just begun our search for natural color source. It is estimated that worldwide up to 70% of all plants have not been investigated fully and that only 0.5% has been exhaustively studied.

REFERENCE

- 1 Praksh, A., Rigelhof, F . and Miller, E. Medallion laboratories analytical progress: Antioxidant activity *Int.J. DeVrie.*, 2001; 1-6.
- 2 Manimala., and R Murugesan., Studies on carotenoid pigment production by yeast *Rhodotorulamucilaginosa* using cheap materials of agro-industrial origin *The Pharma Innovation Journal* 2017; 6(1): 80-82
- 3 Gomez LCM, Montanez J, Zavala AM, Aguilar CN. Biotechnological production of carotenoids by yeasts: an overview. *Microb Cell Fac.* 2014; 13:12.
- 4 Srikalanlayanukul, M., Chartchai, K., Decolorization of Textile wastewater by immobilized *Coriolus versicolor RC3* in Repeated-Batch System with the effect of Sugar Addition. *CMU Jornal.* 2006; 5(3): 301-306
- 5 Gurav, A., J., and Girish, K., Decolourization of anthroquinone based dye Vat Red 10 by *Pseudomonas desmolyticum* NCIM2112. *International Journal for Biotechnology and Molecular Biology.* 2011; 2(6): 93-97.
- 6 Ramendra Singh Parmar*, Charu Singh, Pragya Jadon, Gaurav Bhadauriya and Ajay Kumar Dept. of Life Sciences, ITM University, Gwalior, "Exploration of Pigment Producing Actinomycetes, Isolated From Madhya Pradesh Region of India", *IJHER*, 2016; 6(2), 2016; 13.

- 7 Golubev, W.I. Perfect state of *Rhodomycesdendrorhous* (*Phaffiarhodozyma*). *Yeast*, 1995; 11: 101-110.
- 8 Nagpal, N., Munjal, N. and Chatterjee, S. Microbial Pigments with Health Benefits - A Mini Review. *Trends Biosci*, 2011; 4: 157-160.
- 9 Zheng Zhang, Scott Schwartz, Lukas Wagner, and Webb Miller. "A greedy algorithm for aligning DNA sequences", *J Comput Biol* 2000;7(1-2):203-14.